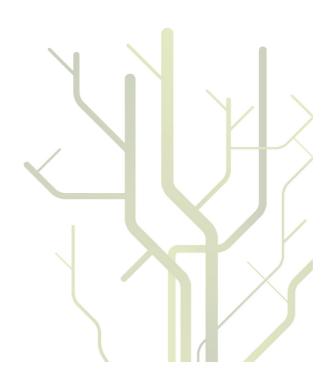


Can vitamin D₃ supplementation prevent bone loss in persons with MS?



Linn Hofsøy Steffensen

A dissertation for the degree of Philosophiae Doctor

Can vitamin D₃ supplementation prevent bone loss in persons with MS?

by

Linn Hofsøy Steffensen

A dissertation for the degree of Philosophiae Doctor

University of Tromsø

Faculty of Health Science

Department of Clinical Medicine

Contents

A(cknowledge	ements	6
No	orsk samme	endrag- Norwegian summary	7
Er	ıglish sumı	mary	9
List of papers			11
Abbrevation			
1.	Introduct	tion	14
	1.1. Multi	ple sclerosis	14
	1.1.1.	Introduction	14
	1.1.2.	Epidemiology of MS	14
	1.1.3.	Classification and clinical manifestations of MS	14
	1.1.4.	Pathogenesis of MS	15
	1.1.5.	Aetiology and risk factors for MS	17
	1.1	1.5.1.Epstein Barr virus	18
	1.1	1.5.2.Sun exposure and vitamin D	18
	1.1	1.5.3.Genetic influence	20
	1.1	1.5.4.Other	21
	1.1.6.	Diagnosing MS	21
	1.1.7.	MS treatment	22
	1.1.8.	The clinical course of MS	22
	1.1.9.	Vitamin D status in persons with MS and the effect on the	
		clinical activity	23
	1.2. Osteo	pporosis	26
	1.2.1.	Bone physiology	26
	1.2.2.	Osteoporosis	27
	1.2.3.	Definition of low bone mass and osteoporosis	29
	1.2.4.	Prevention and treatment of osteoporosis and fracture	30
	1.3. Vitan	nin D	30
	1.3.1.	Vitamin D sources	30
	1.3.2.	Vitamin D metabolism	31
	1.3.3.	Vitamin D effects	33
	1.3.4.	Vitamin D levels and intake	34
	1.4. Multi	ple sclerosis, osteoporosis and vitamin D	35

	1.4.1. Shared pathogenic and aetiological factors for MS and osteoporosis	35
	1.4.2. BMD and osteoporosis in persons with MS	36
	1.4.3. Fracture risk in persons with MS	37
2.	Aims of the study	38
3.	Methods and material	
	3.1. Trial design	39
	3.1.1. Participants	39
	3.1.2. Exclusions criteria	39
	3.1.3. Ethics	39
	3.2. Randomisation and intervention	40
	3.3. Adherence and safety	
	3.4. Clinical investigation	40
	3.5. BMD measurements	41
	3.6. Blood tests	43
	3.7. Questionaires	43
	3.8. Statistical analysis	43
	3.8.1. Statistical tests applied in paper I	44
	3.8.2. Statistical tests applied in paper II	44
	3.8.3. Statistical tests applied in paper III	45
4.	Summary of the results	45
	4.1. Paper I and paper I erratum	45
	4.2. Paper II	46
	4.3. Paper III	47
5.	Discussion	48
	5.1. General discussion	48
	5.1.1. Paper I	49
	5.1.2. Paper II	51
	5.1.3. Paper III	52
	5.2. Methodological aspects of the study	54
	5.2.1. Bias and confounding	54
	5.2.2. Random errors	55
	5.2.3. External validity	55
6.	Implications for further research	56
	6.1. Bone loss and vitamin D supplementation	56

	6.2. Vitamin D levels	56		
7.	Concluding remarks	57		
8.	References	58		
Paper I-III				

Appendices A-C

Acknowledgements

First of all I want to express my sincere thanks to all the participants in this intervention study for their generous contribution.

I want to thank my main supervisor Margitta Kampman for letting me be a part of this interesting project. Thank you for giving me the opportunity to work independently and for always having time to discuss my work when I needed to. I have gained an enormous amount of knowledge. Thank you so much for your patience with me. Thank you, Lone Jørgensen, for being my co-supervisor and contributing with your broad experience.

Magrit Brustad, at the Institute of Community Medicine, thank you for sharing your knowledge about vitamin D with me. Thank you, Svein Ivar Mellgren at the Department of Neurology, for being always positive and help full. I will thank the study nurses at the Department of Neurology, Grethe Berg Johnsen and Marianne Røst, you are two wonderful persons and it is a pleasure working with you. Thanks to the staffs at the Clinical Trial Unit and Inger Sperstad at the Clinical Research Department, you have been very helpful in the planning and performance of this clinical trial. Thank you, Bjørn Straume at the Institute of Community Medicine, you have steadily guided me through the statistical analysis.

Thank you, my colleagues at the department of Neurology who contribute to an including and academic working environment. During these years when I have been working with my dissertation, I am very grateful to have had the opportunity to be involved in the clinical work at our department.

Thanks to the University of Tromsø, the Research Council of Norway and Odd Fellows Research Foundation for Multiple Sclerosis which the study has received financial support from.

Finally, I want to thank all my wonderful family, in particular my husband Rudi and our children, Kamille and Marie, for being so supportive during these years. I love you all so much!

Linn Hofsøy Steffensen Tromsø, September 2012

Norsk sammendrag- Norwegian summary

Multippel sklerose (MS), en betennelsessykdom i nervesystemet og en av de hyppigste årsakene til kronisk nevrologisk funksjonsnedsettelse hos unge i den vestlige verden, er også en mulig årsak til beinskjørhet. Lav beintetthet er hyppigere hos personer med MS enn hos friske personer og flere studier har vist at beintetthet hos personer med MS reduseres etter hvert som funksjonsnedsettelse hemmer fysisk aktivitet. Beintetthet er i hovedsak bestemt av genetiske og hormonelle faktorer, kropps masse indeks, fysisk aktivitet og inntak av kalsium og vitamin D.

Det amerikanske og kanadiske "Insitute of Medicine" og det norske Helsedirektoratet har definert vitamin D nivå i blod ≥ 50 nmol/l som tilstrekkelig, mens flere eksperter anbefaler ≥ 75 nmol/l med tanke på beinhelse. Hovedkildene for vitamin D er sollys, fet fisk og berikede matvarer som margarin og melk. I tillegg finnes det som kosttilskudd. De amerikanske og kanadiske anbefalingene er inntak på 600 IU vitamin D daglig for de mellom 1 år og 70 år. De norske anbefalingene er inntak av 300 IU vitamin D daglig hos personer mellom 2 år og 60 år.

Studier utført hos personer med MS har påvist gjennomsnittlig vitamin D nivå i blod fra omtrent 40 nmol/l til 80 nmol/l. Vitamin D mangel kan bidra til dårlig beinhelse allerede før funksjonsnedsettelsen hemmer fysisk aktivitet. Vi ønsket å se om en ukentlig dose med høydose vitamin D₃ (20.000 IU) tatt i 96 uker kunne hindre beintap hos gangføre personer med MS i alderen 18 til 50 år. 71 personer med MS fikk enten høydose vitamin D₃ eller placebo ukentlig. 68 ble inkludert i analysen. Ved studiestart om vinteren var gjennomsnittlig vitamin D nivå i blod 56 nmol/l og halvparten av deltagerne hadde tilstrekkelige nivåer (≥ 50 nmol/l). Soleksponering i form av solferie eller solarium og jevnlig vitamin D inntak økte sjansen for å ha adekvate verdier. Andelen av deltagerne i placebogruppa som hadde tilstrekkelig nivå økte fra 55 % om vinteren til 92 % om sommeren. Alle deltagerne i gruppa som fikk vitamin D tilskudd oppnådde tilstrekkelig vitamin D nivåer om vinteren, og 91 % oppnådde 75 nmol/l eller høyere.

Ved studiestart hadde 25 % av deltagerne lavere enn forventet beintetthet (z-score ≤ -2.0). Der var ingen sikker forskjell i endring i beintetthet fra studiestart til studieslutt mellom behandlingsgruppene, heller ikke etter justering for vitamin D nivå i blod ved studiestart eller forandring i vitamin D nivå i løpet av studieperioden. I hoftene var der i hele gruppen et gjennomsnittlig beintap på 1 %.

Resultatene fra denne studien med unge gangføre personer med MS kan tyde på at både kvinner og menn med MS har tidlig forhøyet risiko for å utvikle lav beintetthet. Økt funksjonsnedsettelse vil videre kunne bidra til utvikling av beinskjørhet, samtidig som risikoen for fall med påfølgende fraktur tiltar. Det kan være hensiktsmessig å utføre beintetthetsmålinger på alle MS pasienter, mulig like etter diagnosen, men i alle fall når der oppstår påvirkning av gangfunksjonen. Selv om denne studien ikke kunne vise at vitamin D hindret utvikling av beintap, så er vitamin D inntak i anbefalte doser et viktig forebyggende tiltak mot beinskjørhet. Halvparten av deltagerne hadde vitamin D mangel i løpet av vinteren. Alle MS pasienter som i løpet av vintermånedene ikke får soleksponering og som har lavt vitamin D inntak bør bli anbefalt å ta vitamin D tilskudd for å unngå vitamin D mangel.

English summary

Multiple sclerosis (MS), a neuroinflammatory disease and one the most common causes of chronic neurological dysfunction in young adults in the Western world, is also a possible cause of secondary osteoporosis. Several studies have shown that in persons with MS, bone mineral density (BMD) decreases as disability increasingly limits physical activity. In persons with MS low BMD is more prevalent when compared with healthy controls. BMD is primarily determined by genetic and hormonal factors, body mass index, physical activity and intake of calcium and vitamin D.

Serum vitamin D(25[OH]D) level \geq 50 nmol/L is classified as sufficient by the US and Canadian Institute of Medicine and the Norwegian Directorate of Health, however, optimal serum 25(OH)D level for bone health is according to many experts at least 75 nmol/L. The main source of vitamin D for most people is vitamin D generated in the skin during sun exposure. The dietary intake of vitamin D is usually limited, but vitamin D is especially found in fatty fishes and may also be obtained by fortified foods and in supplements. The recommended dietary intake of vitamin D is by the Institute of Medicine 600 IU daily for ages 1-70 years and the Norwegian recommendation is 300 IU daily for ages 2-60 years.

Studies performed in persons with MS have reported mean serum 25(OH)D values ranging from approximately 40 nmol/L to 80 nmol/L. Low vitamin D levels may contribute to poor bone health, even before disability limits physical activity. In this 96 week randomized controlled clinical trial, we aimed to asses whether a weekly dose of 20,000 IU vitamin D_3 could prevent bone loss in fully ambulatory persons with MS age 18-50 years. 71 persons with MS where randomized to receive either a weekly dose of 20,000 IU vitamin D_3 or placebo. 68 were included in the final analysis. At the study start in winter, mean serum 25(OH)D level was 56 nmol/L, and half of the participants had sufficient levels ($\geq 50 \text{ nmol/l}$). Serum vitamin D levels were strongly predicted by sun exposure through sun vacation or solarium and by vitamin D intake. In the placebo group, the proportion of study participants with sufficient levels of vitamin D increased from 55% during winter to 92% during summer. All participants in the intervention group achieved sufficient winter levels. Winter levels of 75 nmol/L or higher were achieved by 91% of the participants.

BMD was lower than expected (z-score \leq -2.0) in 25% of the participants at screening. Percentage change in BMD from baseline to study end did not differ between participants

who had received vitamin D₃ supplementation and those who had received placebo. This result was not altered by adjustment for baseline serum 25(OH)D level or change in serum vitamin D over the study period. However, we found that mean hip BMD decreased significantly in the whole study population by 1.0%.

Findings in our limited sample of young ambulatory persons with MS indicate that men as well as women with MS are at increased risk of low BMD. The results strengthen the suspicion that MS may be a cause of secondary osteoporosis. Progressing physical handicap poses persons with MS at further risk of developing osteoporosis and at the same time increases the risk of falls and fractures. It might be appropriate to perform BMD measurements in all persons with MS, perhaps shortly after diagnosis, but at least when the disease affect the ambulatory function. Even though this small study could not prove any effect of vitamin D supplementation on BMD loss, intake of vitamin D is an important factor in preventing osteoporosis. Half of the participants in this study were vitamin D insufficient during winter months. In order to achieve adequate vitamin D levels, MS patients who have no vitamin D efficient sun exposure and low dietary vitamin D intake during the winter months should be recommended to take vitamin D supplements.

List of papers

Paper I

Steffensen LH, Mellgren SI, Kampman MT (2010) Predictors and prevalence of low bone mineral density in fully ambulatory persons with multiple sclerosis. *J Neurol* 257:410-418 Steffensen LH, Mellgren SI, Kampman MT (2010) Erratum to: Predictors and prevalence of low bone mineral density in fully ambulatory persons with multiple sclerosis. *J Neurol* 257(3):497-498

Paper II

Steffensen LH, Jorgensen L, Straume B, Mellgren SI, Kampman MT (2011) Can vitamin D supplementation prevent bone loss in persons with MS? A placebo-controlled trial. *J Neurol* 258:1624-1631

Paper III

Steffensen L.H, Brustad M, Kampman M.T. (2012) What is needed to keep persons with multiple sclerosis vitamin D-sufficient throughout the year? J. Neurol. Doi: 10.1007/s00415-012-6611-6

Abbrevation

APC: Antigen presenting cell

BBB: Blood brain barrier

BMD: Bone mineral density

BMI: Body Mass Index

CFS: Cerebrospinal fluid

CIS: Clinically isolated syndrom

CNS: Central nervous system

CPBA: Competitive protein binding assay

DBP: Vitamin D binding protein

DMTs: Disease modifying treatments

DXA: Dual X-ray absorptiomatry

EAE: Experimental autoimmune encephalomyelitis

EBV: Epstein Barr Virus

EDSS: Expanded Disability Status Scale

FGF-23: Fibroblast growth factor -23

FS: Functional System

Gd: Gadolinium

HLA: Human leukocyte antigen

HPLC: High-performance liquid chromatography

iCa: Ionized calcium

INF-β: Interferon-β

IRR: Incidence rate ratio

IU: International Unit

LRP5: Low-density lipoprotein receptor-related protein 5

LS/MS: Liquid chromatography coupled with mass spectometry

MBP: Myelin basic protein

MHC: Major histocompatobility complex

MRI: Magnetic resonance imaging

MS: Multiple sclerosis

MSFC: Multiple Sclerosis Functional Composite

OPG: Osteoprotegerin

PBA: Protein binding assay

PPMS: Primary progressive multiple sclerosis

PTH: Parathyroid hormone

RANK/ RANKL: receptor activator of nuclear factor-kappa B/ RANK ligand

RCT: Randomized controlled trial

RIA: Radioimmunoassay

RRMS: Relapsing remitting multiple sclerosis

SEP: Somatosensory evoked potential

SPMS: Secondary progressive multiple sclerosis

TBBM: Total body bone mineral

TGF: Transforming growth factor

UD: Ultradistal

UVR: Ultraviolet radiation

VDR: Vitamin D receptor

VDRE: Vitamin D responsive element

VEP: Visual evoked potential

WHO: World Health Organization

1,25(OH)2D: 1,25-dihydroxyvitamin D

25(OH)D: 25-hydroxyvitamin D

1. Introduction

1.1 Multiple Sclerosis

1.1.1 Introduction

Multiple sclerosis (MS) is one the most common cause of chronic neurological dysfunction in young adults in the Western world. In MS, inflammation and chronic degeneration leads to demyelination and axonal destruction in the central nervous system (CNS) and it is a multifactorial disorder, believed to be a result of both genetical and environmental factors and has a female predominance [Compston and Coles 2008].

1.1.2 Epidemiology MS

The estimated worldwide prevalence of MS is between 1-1 and 2.5 million cases [Miller 2012]. Norway is a high risk area of MS, with the lowest prevalence in the northernmost counties Finnmark and Troms (86-104 per 100.000) and the highest in the southernmost part (180-190 per 100.000) [Kampman *et al.* 2007; Vatne *et al.* 2011].

1.1.3 Classification and clinical manifestations of MS

The majority of the patients (80-90 %) present during their second or third decade of life with the relapsing remitting form of MS (RRMS) [Noseworthy et al. 2000; Tremlett et al. 2010] which is characterised by clinical exacerbations of neurological symptoms, followed by complete or incomplete remission [Miller 2012]. After 10-20 years about 65% -70% of the RRMS patients enter a secondary progressive phase, secondary progressive MS (SPMS) [Compston and Coles 2008]. The remaining 10-20% of the patients experience a primary progressive form of MS (PPMS) with progressive clinical deterioration from the onset of the disease, without the characteristic exacerbations. Age at onset is around 40 years or later and there is a male predominance [Miller and Leary 2007; Compston and Coles 2008]. The patients with RRMS present with an acute episode affecting one (or occasionally several) sites in CNS which develops rapidly and gradually improves after days to weeks [Compston and Coles 2008]. The symptoms and signs generally reflect those parts of the CNS where there is a dense concentration of myelinated fibres subserving motor and sensory functions [Compston et al. 2006]. The first appearance are typically sensory disturbances, unilateral optic neuritis, diplopia (internuclear ophtalmoplegia), Lhermitte's sign (trunk and limb parasthesias evoked by neck flexions), limb weekness, clumsiness, gait ataxia and neurogenic

bladder and bowel syndrome. In PPMS the disease is progressive from the onset [Compston and Coles 2008]. The most common presentation is progressive spastic paraparesis with an increasing functional impairment over months to years which do not reverse. Excessive fatigue is a common complaint in all subtypes of multiple sclerosis [Compston et al. 2006] and patients with MS may experience both nociceptive pain due to reduced mobility, postural changes and spasticity, and neuropathic pain due to lesion affecting the somatosensory system. Affective disturbance and cognitive deficits are also common in MS patients [Compston et al. 2006]. The patients may experience progressive impaired disability due to limb weakness, spasticity, ataxia, and tremor [Compston et al 2006]. Disability in multiple sclerosis is quantified with The Kurtzke Expanded Disability Status Scale (EDSS) from 0, with a normal neurological examination to 10, death due to MS [Kurtzke 1983]. The EDSS quantifies disability in eight Functional Systems (FS) [Fischer et al. 1999]: mental, visual, sensory, brainstem, pyramidal, cerebellar, bowel and bladder functions and other (includes any other neurological findings due to MS). In addition to FS score the EDSS system is based on the patient's ability to walk. With a walking distance below 500 meters, the EDSS is graded 4.5 or higher. If the patient is in need of assistance to walk (cane, cruch or brace) the EDSS is graded 6.0 or higher. If the patients is restricted to bed or chair the EDSS is graded 8.0 or higher.

1.1.4 Pathogenesis of MS

The hallmark of MS pathology is formation of the sclerotic plaque in the CNS which represents the end stage of a process involving inflammation, demyelination and remyelination, oligodendrocyte depletion and astrocytosis, and neuronal and axon degeneration [Compston and Coles 2008]. MS is considered a T-cell disease involving activated myelin-specific auto reactive T cells [Frohman *et al.* 2006]. Myelin is a protein which is synthesised by mature oligodendrocytes in the CNS. It is extending along the nerve fibres in the white matter tract and is necessary to properly propagate nerve impulses between neurons [Frohman *et al.* 2006]. The immunepathogenesis, shown in figure 1, may be described in different phases [Miller 2012]. First myelin specific reactive T cells are activated outside the CNS via a process in which cell surface receptors recognize processed antigens in association with major histocompatobility complex (MHC) class II molecules (also called human leukocyte antigen [HLA]) on antigen presenting cells (APCs). Then they become activated through one of several costimulatory molecules, such as B7/CD28 or CD40/CD40L. How these auto reactive T-cells are activated in the periphery is still a matter of debate.

Processes like molecular mimicry, where T-cells generated against non-self (virus or microbial antigens) cross-react with self-myelin and T cell activation triggered by myelin antigens presented in lymph nodes have been postulated as potential mechanism [Holmoy 2007; Comabella and Khoury 2012]. These activated T cells can proliferate and differentiate into one of several subtypes of T helper (Th) cells (also called CD4+ cells) which secrete cytokines to facilitate different types of immune response. Th1 cells are considered proinflammatory and produce proinflammatory cytokines, opposed to Th2 cells that secret anti-inflammatory cytokines. A reduction in the effector functions of regulatory T cell has also been observed [Viglietta et al. 2004]. After activation in the periphery the T cells adhesions molecules bind to and cross the blood brain barrier (BBB) [Dhib-Jalbut 2007]. Within the CNS the myelin-spesific CD4+ T cells may be reactivated in situ by myelin antigens presented on MHC II molecules on antigen presenting cells (APC), such as microglia or macrophages, and trigger release of proinflammatory cytokines which further disrupt the BBB and stimulate chemotaxis with a second larger wave of inflammatory cells to the CNS [Comabella and Khoury 2012]. After re-activation T cells proliferate and secrete proinnflammatory cytokines which stimulate microglia, macrophages, astrocytes and recruited B cells and ultimately result in demyelisation.

The most studied animal model of MS is the experimental autoimmune encephalomyelitis (EAE). In 1981 Ben-Nun et al isolated myelin basic protein (MBP)-specific CD4+ T cells from rats immunized with purified MBP and cultured them in vitro. The T line cells were then injected in healthy recipients from the same rat strain that had been immunised and after 4-5 days they developed experimental autoimmune encephalomyelitis (EAE) [Ben-Nun *et al.* 1981]. EAE may also be induced by immunizing animals with other myelin derived protein or peptides [Comabella and Khoury 2012]. The EAE has exerted a major influence on MS research, although it fails to reflect some areas in the MS pathogenesis; the B-cell component and intrathecal production of IgG, how the immune-system breaks the self-tolerance and the relation between the neuroinflammation and neurodegeneration [Holmoy 2007].

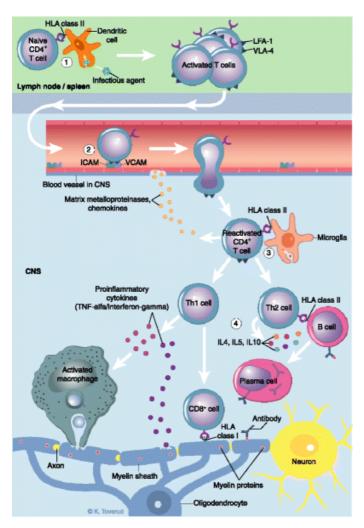


Fig. 1: The MS immunepathogenesis. Reproduced with permission from [Holmoy 2007]

1.1.5 Aetiology and risk factors for MS

The aetiology of MS remains elusive, but it is assumed that both environmental factors and a complex genetic background contribute to the disease [Comabella and Khoury 2012], the first probably accounting for as much as 80 % of the risk [Ebers 2008]. The proposed and most extensively studied environmental triggers are Epstein Barr virus, low sunlight, low vitamin D, smoking, diet and toxins [Compston and Coles 2008;Pugliatti *et al.* 2008a]. Migration of people affect the distribution of MS, and migration in childhood from a high-risk to a low-risk region is associated with a reduced MS risk and from a low-risk to a high-risk is associated with a higher MS risk. The age-at-migration was originally suggested to be before 15 years of age, but later studies from Australia found that the risk of environmental factors in multiple sclerosis may operate over a period of many years and not only in childhood and early life [Hammond *et al.* 2000]. Genetic epidemiologic studies strongly indicate that environmental

factors act at a broad population level, rather than in the familial microenvironment [Orton *et al.* 2008].

1.1.5.1 Epstein Barr virus

Several microbes have been suggested to be associated with the risk of MS, but the evidence linking MS and Epstein Barr virus (EBV) are particularly strong. EBV infection appears to be a necessary (but not sufficient) condition for adult MS to develop [Holmoy 2007] and the risk is highest among individuals first infected with EBV in adolescence or later in life, often manifested as infectious mononucleosis [Thacker *et al.* 2006].

1.1.5.2 Sun exposure and vitamin D

One of the most striking features of MS is the geographic distribution with a positive latitude gradient, showing, with some notable exceptions, a pattern of high MS frequency in areas where sunlight exposure is low and, consequently, opportunity for vitamin D synthesis in the skin is limited [Pugliatti et al. 2006; van der Mei et al. 2001; Kampman and Brustad 2008;Sloka et al. 2011;Vukusic et al. 2007;Ascherio and Munger 2007;Compston and Coles 2008] However only one prospective study so far has shown that circulating levels of $25(OH)D \ge 100 \text{ nmol/L}$ were associated with a lower risk of MS in whites [Munger et al. 2006].Later findings have suggested an attenuation of the latitude gradient [Alonso and Hernan 2008; Pugliatti et al. 2002], but MS prevalence in Australia and in France could be closely predicted by regional UVR levels [van der Mei et al. 2001; Vukusic et al. 2007], and a geospatial analysis confirmed a strong association between UVR and MS distribution [Beretich and Beretich 2009]. A recently published large case-control study from Sweden found a significant inverse relationship between exposure to ultraviolet radiation (UVR) and risk of developing MS [Baarnhielm et al. 2012]. Ecological studies have also shown an inverse correlation of sunlight exposure through outdoor activities during the period spanning from childhood to adolescence, but also into adulthood, with MS risk [Hammond et al. 2000; Freedman et al. 2000; van der Mei et al. 2003; Goldacre et al. 2004], even north of the Arctic Circle [Kampman et al. 2007]. Possible pathways for immunosuppressive effects of UVR exposure in immune-mediated disorders that are likely to be independent of vitamin D synthesis are also recognised [Beretich and Beretich 2009; Lucas and Ponsonby 2006].

The effect of dietary vitamin D on MS risk has been less studied than the correlation with UVR. Most humans are not dependent on dietary vitamin D supplies [Hollis 2005], and a

correlation of vitamin D from food with 25(OH)D is only found when UVR exposure is low [Brustad *et al.* 2004;van der Mei *et al.* 2007;Macdonald *et al.* 2008] In the Nurses' Health Study following 187,000 women aged 25 to 55 years at inclusion, total vitamin D intake and use of vitamin D supplements ≥ 400 IU/day, but not vitamin D intake from food, were associated with higher serum 25(OH)D levels and lower MS risk [Munger *et al.* 2004]. However, it must be commented that the inclusion age was quite high given that median age of onset of MS is 30 years and that environmental factors may act many years before disease onset. In addition, both vitamin D intake from food and supplement was rather low. In a MS case-control study in Norway north of the Arctic Circle, use of cod-liver oil supplements when growing up was associated with less MS in the subgroup of respondents with low summer outdoor activities [Kampman *et al.* 2007]. In a case-control study from Sweden, intake of fatty fish and vitamin D supplement use had only minor influence according to the risk of MS development [Baarnhielm *et al.* 2012].

Animal studies of EAE have shown that induction of EAE could be prevented and EAE progression blocked by whole body UV –irradiation and that vitamin D hormone supplementation completely inhibited EAE induction and progression [Hauser *et al.* 1984;Cantorna *et al.* 1996;Lemire and Archer 1991]

A role of vitamin D in the immunopathogenesis of MS is biologically plausible. Vitamin D is a potent immune modulator [Adams and Hewison 2008;Fernandes de Abreu *et al.* 2009], and a role of vitamin D has also been proposed in other immune-mediated diseases, a.o. diabetes type I, rheumatoid arthritis, and inflammatory bowel disease [Fernandes de Abreu *et al.* 2009]. Proliferation assays show an association of high 25(OH)D levels with an improved regulatory T cell function in persons with MS [Correale *et al.* 2009;Smolders *et al.* 2009b]. Also significant correlations between vitamin D status and immunological markers have been reported in cross sectional studies, which might indicate an effect on the disease activity by vitamin D, however these correlations might be caused by an effect of the disease on the vitamin D status [Ascherio *et al.* 2010]. Direct genomic signalling by active 1,25(OH)₂D occurs through the vitamin D receptor (VDR), which is present in multiple cells of the immune system as well as in neurons and glial cells in the human brain [Smolders *et al.* 2008a;Eyles *et al.* 2005]. Activation of the VDR by hormonal vitamin D stimulates a shift from proinflammatory Th1 responses to anti-inflammatory Th2 responses [Smolders *et al.* 2008a]. Holmøy was the first to propose that poor vitamin D status modulates the immune

response to EBV in a way that increases the risk of developing MS [Holmoy 2008;Hayes and Donald 2008].

1.1.5.3 Genetic influences

Family studies have revealed that first –degree relatives of a person with MS are generally at 15-35 times greater risk of developing MS compared with the general population and the risk correlates with the degree of kinship [Ramagopalan et al. 2009]. There are several genes that seem to be associated with MS. The association between MS and alleles of the MHC (HLA) classes I and II was identified in the early 1970s [Compston and Coles 2008], but variants so far identified explain only approximately 50 % of the inherited risk of MS [Ramagopalan et al 2009]. T-cell receptor, CTLA4 and ICAM1 are other genes associated with MS [Dyment et al. 2004]. Reports on novel gene-environment interactions continue to increase our understanding of the role of vitamin D in MS. The HLA-DRB1*1501 risk haplotype is a strong genetic predictor of MS risk and a vitamin D responsive element (VDER) has been identified in the promoter region of this haplotype [Ascherio et al. 2010;Ramagopalan et al. 2009]. Conversely, a significant genetic influence on regulation of circulating 25(OH)D concentrations has been found in MS twins [Orton et al. 2008]. The increasing ratio in the concordance of MS risk between monozygotic and dezygotic twins with increasing latitude also suggests that genetic effects may be stronger at lower UVR exposure and/or lower vitamin D status [Islam et al. 2007]. However, still little is known about the role of vitamin D related genes or specific genetic interactions with vitamin D in determining MS risk [Ascherio et al. 2010] and the contribution of VDR gene polymorphisms to immune regulation in MS is not fully understood [Smolders et al. 2009a]. First evidence has been provided that a functional variant of the VDR gene interacts with sun exposure in childhood to influence MS risk [Dickinson et al. 2009], implying that vulnerability to poor vitamin D status may be determined by genetic variations. The modification of the association of past sun exposure with MS risk by "red hair colour" genotype provides further support for a causal effect of UVR/vitamin D in the aetiology of MS [Dwyer et al. 2008]. The disproportional increase in the incidence of MS in women that was first observed in Canada is likely to be caused by sexspecific exposure or susceptibility to environmental factors [Orton et al. 2006]. Data supporting an interaction between female sex, possibly mediated by oestrogen, and vitamin D in MS risk are accumulating. Protective effects of sun exposure are mainly observed in female MS patients [Islam et al. 2006; Woolmore et al. 2007; Dwyer et al. 2008; Kampman and Brustad 2008]. Sex differences in vitamin D metabolism were first reported in the EAE

model: A cholecalciferol containing diet inhibited severe EAE only in female mice, indicating a sex difference in vitamin D metabolism in the CNS [Spach and Hayes 2005]. The same group showed recently that 17β-estradiol is essential for VDR gene expression and function in the inflamed CNS in EAE mice [Nashold *et al.* 2009]. Stronger immunomodulatory effects of vitamin D on CD4 T cells and an increase in the number of T regulatory cell in women than in men have been shown [Correale *et al.* 2010]

1.1.5.4 Other

Several studies have described an association between smoking and MS risk, and although confounding cannot be excluded, the evidence suggests that smoking increases the risk for MS and also likely accelerates the progression of the disease [Ascherio and Munger 2007]. A possible association between toxins and MS risk and especially on occupational exposure to organic solvents has been proposed, but the results are divergent [Marrie 2004;Pugliatti *et al.* 2008a]. Estrogen in high levels appear to shift the immune response from the proinflammatory type I, to the noninflammatory type II and may be associated with reduced risk of MS [Ascherio and Munger 2007]. Evidence remains mostly thin, but diets high in saturated fat/low unsaturated fat and low in dietary antioxidant may influence MS risk [Ascherio and Munger 2007].

1.1.6 Diagnosing MS

MS is a clinical diagnosis, and the medical history and physical examination are the most important part in diagnosing the disease. However, Magnet Resonance Imaging (MRI) of the CNS is playing a significant role in the clinical and scientific investigation of MS. In at least 95 % of the patients conventional MRI typically show the characteristic demyelinated plaque, described as a T2 lesion in the white matter as a result of tissue damage [Compston *et al.* 2006]. During acute exacerbations the local increase of BBB permeability can be imaged using MRI after the administration of a contrast material as gadolinium (Gd) [Bruck *et al.* 1997]. Cerebrospinal fluid (CSF) analysis often shows intrathecal synthesis of immunglobulins, presented as oligoclonal band or increased IgG production and moderate pleocytosis. Positive CSF finding are defined as either elevated IgG index (in relation to serum) or two or more oliclonal bands [Polman *et al.* 2011], seen in 90 % of the MS patients [Compston and Coles 2008]. Dysfunctions of the optic nerve and spinal cord may be demonstrated by visual evoked responses (VEP) and somatosensory evoked potential (SEP) [Noseworthy *et al.* 2000].

According to the McDonald Criteria [Polman *et al.* 2011] for patients with two or more attacks who have objective clinical evidence of two or more lesions or objective clinical evidence of one lesion with reasonable historical evidence of a prior attack, no additional data are required. The criteria for diagnosis of RRMS are two or more clinical exacerbations with either objective clinical evidence or MRI showing the typically changes, or one clinical exacerbation with objective clinical evidence and MRI showing both T2 lesions and GD enhanced lesions [Polman *et al.* 2011]. Clinically isolated syndrome (CIS) is diagnosed in patients with one clinical exacerbation not fulfilling the MRI criteria for the MS diagnosis. The criteria for diagnosis of PPMS are one year of disease progression, and two ore more of the following findings; Both T2 and Gd enhaced lesions on MR of the brain, two or more T2 lesions in the spinal cord and positive CSF findings [Polman *et al.* 2011].

1.1.7 MS treatment

The medical treatment of MS consists of treatment of the acute exacerbations and several disease modifying therapies (DMTs). Acute exacerbations with moderate to serious disability are treated with high dose short-term methylprednisolone for 3-5 days to speed up the recovery, but the treatment has no influence on the occurrence of new relapses or long-term disability [Myhr and Mellgren 2009]. The goals of the DMTs are decrease the relapse rate, to arrest or slow the progression of disability and subclinical disease progression on MRI and to maintain or improve quality of life. The first line treatment consists of Interferon-β (INF-β) and Glatiramer acetate. The second line treatment consists at this moment of two agents, the Natalizumab and Fingolimod [Polman *et al.* 2006;Pelletier and Hafler 2012;Kappos *et al.* 2010]. A third line treatment with chemotherapy, mitoxantrone, is also possible. In addition several symptomatic treatments are available against pain, spasticity, mood changes, urinary/bowel dysfunction and walking disabilities [de Sa *et al.* 2011].

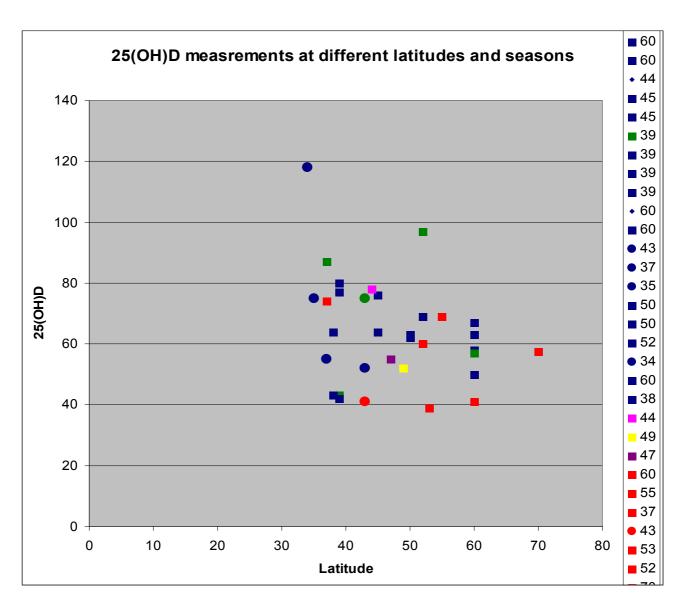
1.1.8 The clinical course of MS

Several factors have been proposed modifying the disease progression and exacerbations in MS. The relapsing form of MS is generally associated with a better prognosis and these patients are expected reaching irreversible disability later than those with a progressive disease [Confavreux *et al.* 2000;Tremlett *et al.* 2006]. MS relapse reduction during pregnancy and a rebound in the puerperium has been found [Confavreux *et al.* 1998] and the explanation has been that estrogens in high levels appear to shift the immune response from the

proinflammatory type I to the noninflammatory type II [Ascherio and Munger 2007]. However it is generally agreed that there is no difference in overall prognosis between women who have been pregnant compared with those who have not. Smoking, in addition to increase the risk of MS, also likely accelerates the progression of the disease [Ascherio and Munger 2007]. Evidence remains mostly thin but diets high in saturated fat/low unsaturated fat and low in dietary antioxidant may also influence MS progression [Ascherio and Munger 2007]. Possible due to proinflammatory responses, studies have revealed a significant association between psychosocial stress and MS exacerbations [Mohr *et al.* 2004]. However, one of the most studied prognostic factors for MS is the vitamin D, described in 1.1.9.

1.1.9 Vitamin D status in persons with MS and the effect on the clinical activity

Vitamin D status probably declines after MS onset [Munger et al. 2006]. Studies that measure 25(OH)D in MS patients are therefore uninformative as to whether higher vitamin D status decreases MS risk [Ascherio et al. 2010]. The only prospective study is a nested casecontrol study among more than 7 million US military personnel that found mean serum 25(OH)D level between 70.3 nmol/L and 73.5 nmol/l before MS onset and 63.3 nmol/L after MS onset [Munger et al. 2006]. Most observational studies comparing 25(OH)D levels in Caucasian MS patients and controls report on less than 50 individuals in each group and the reported mean 25(OH)D levels ranging from approximately 40 to 80 nmol/l [Amezcua et al. 2012; Barnes et al. 2007; Burton et al. 2010; Cosman et al. 1998; Hiremath et al. 2009; Holmoy et al. 2009;Kimball et al. 2007;Knippenberg et al. 2011;Loken-Amsrud et al. 2012;Lonergan et al. 2011; Lucas et al. 2011; Nieves et al. 1994; Orton et al. 2008; Ozgocmen et al. 2005; Pierrot-Deseilligny 2009; Runia et al. 2012; Simpson S Jr et al. 2010; Smolders et al. 2008b; Soilu-Hanninen et al. 2005; Soilu-Hanninen et al. 2008; Stein et al. 2011; van der Mei et al. 2007; Weinstock-Guttman et al. 2011; Yildiz et al. 2011; Triantafyllou et al. 2012;Baarnhielm et al. 2012]. The largest study to date comparing 25(OH)D levels in 1013 incident cases of MS and 1194 matched controls found mean values of 62.9 nmol/L in cases and 66.3 nmol/L in controls, a difference that is unlikely to be of clinical significance [Baarnhielm et al. 2012]. Figure 2 shows measured vitamin D status in relation to latitude and season.



☐ Northern latitude ☐ Southern latitude

Blue: Whole year Red: Winter Green: Summer Pink: June-March Yellow: June-

February Purple: February-April

Fig. 2: Measured vitamin D status in relation to latitude and season.

Associations of 25(OH)D with a.o. disability, body mass index (BMI), and sex limit comparability of results. Studies in Hispanics and African Americans report lower serum 25(OH)D than in whites, which may be explained by darker skin tones and socioeconomic and cultural differences regarding dietary sources of vitamin D [Amezcua *et al.* 2012;Gelfand *et al.* 2011]. Results of studies comparing 25(OH)D in persons with MS and healthy individuals are inconsistent: levels in persons with MS are reported to be lower than [Ozgocmen *et al.* 2005;Correale *et al.* 2009], higher than [Holmoy *et al.* 2009;Hiremath *et al.*

2009] or equal to [Soilu-Hanninen *et al.* 2008;Barnes *et al.* 2007;van der Mei *et al.* 2007] levels in controls. Levels of 25(OH)D have also been measured in CSF and there were no significant differences between CSF 25(OH)D in 36 persons with RRMS compared with persons with other inflammatory or non-inflammatory neurological diseases [Holmoy *et al.* 2009].

Some observational studies have suggested a relationship between low serum vitamin D levels and disease activity in relapsing-remitting MS, but the causality of this relationship has not been proven. Lower vitamin D levels have been reported during relapses than remission in RRMS patients [Correale et al. 2009; Soilu-Hanninen et al. 2005; Soilu-Hanninen et al 2008] and serum levels of 25(OH)D were associated with both relapse rate [Smolders et al. 2008b; Tremlett et al. 2008] and disability [van der Mei et al. 2007; Smolders et al. 2008b] in MS patients. Studies among veterans in the Multiple Sclerosis Surveillance registry have found that among veterans with RRMS low sun-exposure during the ages of 6-15 years was significantly associated with earlier symptom onset and intake of cod-liver at the same age was associated with later onset of MS, and among those with the progressive forms of MS low sun exposure increased the risk of disease progression and intake of cod liver oil delayed the progression [McDowell et al. 2011]. One report showed that children with higher serum 25(OH)D concentrations at presentation with an acquired demyelinating syndrome had a lower risk of early MS diagnosis [Hanwell H. et al. 2009]. Brain MRI parameters are commonly used as surrogate markers for MS disease activity. In patients with relapsing remitting MS, no correlation was found between serum 25(OH)D and MRI parameters [Soilu-Hanninen et al. 2008]. More indirectly, variations in relapse rate, markers of inflammation, and number of brain lesions on MRI have, with few exceptions, shown a seasonal pattern that can be related to variation in UVR exposure and vitamin D status [Tremlett et al. 2008; Balashov et al. 1998; Embry et al. 2000; Killestein et al. 2002; Van Amerongen et al. 2004; Gray O. et al. 2009]. Although the studies from other races found lower serum 25(OH)D than in whites, they did not find that the measurements varied with increasing disability [Amezcua et al. 2012;Gelfand et al. 2011]. It is known that the vitamin D receptor polymorphisms has evolved differently among populations, but how this influences vitamin D status and MS is unknown [Amezcua et al. 2012].

In a pilot study, vitamin D supplementation has been associated with more favourable clinical outcomes [Burton *et al.* 2010], whereas three small randomised controlled trials did not find

any beneficial clinical effects [Stein *et al.* 2011;Kampman *et al.* 2012;Soilu-Hanninen *et al.* 2012]. One of these studies reported a reduction in the number of T1 enhancing lesions on MRI in the high-dose vitamin D group [Soilu-Hanninen *et al.* 2012]. In a 28 week safety study of increasing daily doses of vitamin D (4000 to 40,000 IU cholecalciferol), the overall number of MRI lesions decreased significantly from baseline to the end of the trial [Kimball *et al.* 2007]. Effect of vitamin D supplementation on other surrogate markers of disease activity has also been reported. Supplementation with 1,000 IU cholecalciferol increased serum levels of the anti-inflammatory cytokine transforming growth factor (TGF)-β1 [Mahon *et al.* 2003]. Randomised controlled studies with high dose vitamin D supplementation, with sufficient statistical power for clinical endpoints, have not yet been published.

1.2 Osteoporosis

1.2.1 Bone physiology

Bone strength is a function of both bone density and quality. The bone must be both stiff and flexible enough to resist deformation and fracture, and it undergoes active growth, modelling and remodelling [Bonnick 2004]. The bone tasks, in addition to provide a frame to keep the body supported, are production of blood cells in the bone marrow and it constitutes the body's store of the minerals, calcium and phosphate [Heaney et al. 2000]. The bone matrix consists of type I collagen stiffened by crystals of calcium hydroxyapatite. The bone cells principally consist of three different cells, the osteoblasts which are responsible for the bone matrix formulation, the osteoclasts which are the bone resorption cells, and the most numerous, the osteocytes that are osteoblasts that have been surrounded by the bone matrix that they have synthesised [Seeman and Delmas 2006]. The skeleton is composed of two types of bone [Bonnick 2004]. Cortical bone is compact bone which typically is in the long bones and the surface of flat bones. Trabecular bone is spongious bone which is primarily in the vertebral bodies, pelvis and in the distal ends of long bones, and it is the trabecular bone that contains the bone marrow, either hematopoietic or fat. The peak bone mass is achieved during the third decade [Heaney et al. 2000]. About three-fourths of the variance in peak bone mass is determined by genetic factors, where important involved genes are those related to the body size, (hormones, receptors and proteins involved in the growth hormone/IGF axis), to the sex steroids, to the vitamin D receptor and to several skeletal cytokines [Heaney et al. 2000]. Studies have identified genetic variants that regulate bone mass, including low-density lipoprotein receptor-related protein (LRP5), osteoprotegerin (OPG) and receptor activator of

NK-κβ (RANKL) [Styrkarsdottir *et al.* 2009]. The OPG/RANK pathway and its ligand (RANKL) are the most important mediators of osteoclast activity, where binding of the RANKL to RANK stimulates the activity and binding of OPG prevents RANKL binding to the RANK. LRP5 is the most important mediator of osteoblast activity and bone formation [Sandhu and Hampson 2011]. The balance between OPG/RANKL signalling and the level of active OPG regulate the bone metabolism [Kong *et al.* 2000]. The pathogenesis of osteoporosis following decline in oestrogen levels are thought to be mediated through the OPG/RANKL system [Hearn and Silber 2010]. Three hormones play a significant role in bone development: parathyroid hormone (PTH), calcitonin and vitamin D. PTH speeds up bone breakdown, calcitonin conserves calcium and vitamin D increases calcium absorption in the intestine. The bone mass accretion is also influenced by the nutrition during years of growth, and especially the intake of calcium, vitamin D, phosphorous and protein, exercise, hormonal status and smoking [Heaney *et al.* 2000]. During the fourth or fifth decade an age related bone loss starts, resulting in a progressive decline in the bone mineral density (BMD) [Poole and Compston 2006].

1.2.2 Osteoporosis

Osteoporosis is a systemic skeletal disease characterized by low bone mass and microartitechural detoriation of bone tissue, leading to increase in bone fragility and susceptibility to fractures [Bonnick 2004;Raisz 2005]. Figure 3 shows scanning electron micrographs of the structure of L3 vertebra in a 31 year old woman (top) and in a 70 year old woman (bottom) were many of the plate-like structures have become converted to thin rods. Osteoporosis can occur because of failure to achieve peak bone mass [Sandhu and Hampson 2011;Heaney *et al.* 2000], when there is an excessive bone resorption resulting in decreased bone mass and microarchitectural deterioration, and when there is inadequate formation response to an increased resorption [Raisz 2005;Sandhu and Hampson 2011].

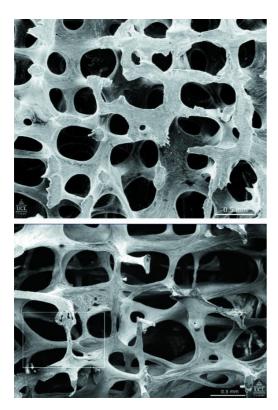


Fig. 3: The picture shows scanning electron micrographs of the structure of L3 vertebra in a 31 year old woman (top) and in a 70 year old woman (bottom). Reproduced with permission from [Poole and Compston 2006]

Figure 4 shows a diagrammatic representation of the bone mass life-line in individuals who achieve their full genetic potential for skeletal mass and those who do not. Oestrogen deficiency is critical to the pathogenesis of osteoporosis in both sexes, but postmenopausal women are at the highest risk for developing the disease [Raisz 2005]. Decreased calcium and/or vitamin D deficiency may result in secondary hyperparathyroidism and reduced mineralisation of the osteoid with low bone mass /osteoporosis as a result. Also immobility and loss of mechanical loading, cigarette smoking and alcohol consume and maternal history of hip fracture are other risk factors and possible reasons to accelerated bone loss and osteoporosis [Raisz 2005;Poole and Compston 2006]. Several underlying diseases (Endocrine, hematologic, rheumatologic and connective tissue diseases and gastrointestinal disorders, Parkinson disease, stroke with hemiplegia and Multiple Sclerosis) may lead to secondary osteoporosis [Hofbauer *et al.* 2010;Poole and Compston 2006;Kampman *et al.* 2011]. Numerous drugs (e.g. glucocorticoids, anticonvulsants, immunosuppressive agents, antidepressants and several hormonal drugs) may lead to drug-induced osteoporosis [Hofbauer *et al.* 2010]. Although low bone mass has a major role in the pathogenesis of

fracture, factors related to falling, risk of falling and protective response make important contributions [Poole and Compston 2006].

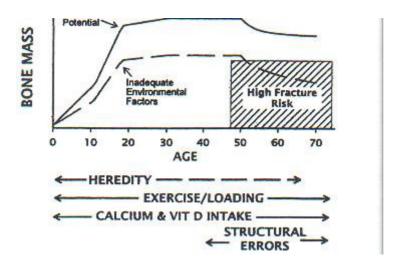


Fig. 4: Diagrammatic representation of the bone mass life-line. Reproduced with permission from [Heaney *et al.* 2000]

1.2.3 Definition of low bone mass and osteoporosis

Diagnosis of osteoporosis is based on measurements of the BMD in the spine and hip, using dual X-ray absorptiometry (DXA) which is considered the gold standard. The examination is non-invasiv and the radiation dose is extremely low. Quantitative ultrasound, based on the measurements of speed of sound is also applied. BMD is reported as a comparison to the peak population mean in sex-matched young healthy adults, the T-score, or a sex-matched and agematched healthy population, the Z-score. The World Health Organization (WHO) has defined osteoporosis as a T-score of less or equal to -2.5 and osteopenia as a T-score between -1.0 and -2.5. The T-score criteria are used for postmenopausal women and men aged 50 years and older [Bonnick 2004; Poole and Compston 2006]. T-scores are designed to predict fracture risk in postmenopausal women. Age is an important predictor of fracture risk. At the same BMD, an older woman is more likely to have a fracture compared with a younger woman of reproductive age, most likely because of lower bone quality [Kanis 2002]. Therefore, T-scores do not predict the same fracture risk in younger women of reproductive age as they do in older, postmenopausal women. A position statement by the International Society for Clinical Densitometry recommends that Z-scores, not T-scores, should be preferred in women before menopause and males under age 50 [Leslie et al. 2006]. A Z-score of \leq 2.0 is "below the expected range for age". Substitution of DXA Z-scores for T-scores gave significant

diagnostic disagreement and significantly fewer persons being diagnosed with low BMD [Carey *et al.* 2009]. For premenopausal women and men younger than 50 years of age the diagnosis of osteoporosis requires in addition to Z-score of \leq 2.0, the presence of a fragility fracture or a recognized risk factor for secondary osteoporosis [Writing group for the ISCD position development conference 2004].

1.2.4 Prevention and treatment of osteoporosis and fractures

The primary risk of osteoporosis is fragility fracture, and the degree of mortality and morbidity related to the fracture are dependent on the localisation and the severity of the fracture. Hip fractures have the most serious impact, with high mortality and morbidity, spinal fractures are associated with pain and loss of height and both cause loss of mobility and autonomy which represent a loss of quality of life [Sambrook and Cooper 2006]. Most fractures will occur in the relatively large group of persons with osteopenia, rather than in the smaller group with osteoporosis, although the risk of fracture is higher in the osteoporosis group [Siris et al. 2004]. Both prevention and the non-pharmacological intervention in treatment of osteopenia and osteoporosis includes general lifestyle factors, such as a balanced diet containing calcium and vitamin D, smoking cessation and avoidance of heavy alcohol use and regular exercise including weight-bearing and muscle strengthening exercise [Sandhu and Hampson 2011]. Exercise programmes incorporating balance, gait and strength training do reduce risk and rate of falls [Gillespie et al. 2009]. The basis of all osteoporosis treatment is supplementation with calcium and vitamin D, in addition pharmacological intervention includes medications that inhibit the bone resorption and stimulate the bone formulation [Poole and Compston 2006]. Oestrogen hormone replacement is regarded as second line therapy because the fracture prevention is outweighed by the higher risk of breast cancer, coronary heart disease, stroke and thromboembolism [Poole and Compston 2006; Sandhu and Hampson 2011]. Evidence does not support treatment with calcium and vitamin D alone, except in institutionalised elderly people and people that have low levels in the blood [Gillespie et al. 2009;Poole and Compston 2006].

1.3 Vitamin D

1.3.1 Vitamin D sources

Vitamin D generally refers to two fat soluble prohormones, vitamin D₂ (ergocalciferol) and vitamin D₃ (cholecalciferol) [O'Mahony *et al.* 2011]. The dietary intake of vitamin D is

usually limited, but vitamin D₃ is found in fatty fishes, raw egg yolk, and vitamin D₂ comes from yeast and plants [O'Mahony et al. 2011; Diehl and Chiu 2010]. Both vitamin D₂ and vitamin D₃ may also be obtained by fortified foods like milk, butter, yogurt and bread [O'Mahony et al. 2011] and by vitamin D supplements, like cod-liver oil. Since vitamin D₂ metabolites make up only a minor part of the total circulating vitamin D content of the body [Heaney et al. 2011], vitamin D₃ is presumably the most relevant. The main source of vitamin D for most people is vitamin D₃ generated by photosynthesis in the skin in which UVR converts cutaneous 7-dehydoxycholesterol into pre-vitamin D₃, which is quickly converted by heat isomerisation to vitamin D₃ [Pearce and Cheetham 2010; Diehl and Chiu 2010]. Cutaneous syntheses of vitamin D depend on pigmentation, age, gender clothing and sunscreen use, in addition to latitude and local weather conditions [Rosen 2011]. The contribution from the different vitamin D sources are in addition to latitude and season dependent of living condition and outdoor activities, genetic traits and culture behaviour [Mithal et al. 2009]. A metanalysis has shown that measured values of 25(OH)D decreased with latitude in Caucasians, but not in non-Caucasians [Hagenau et al. 2009]. Because of seasonal variation of UVR, cutaneous vitamin D production may be absent during part of the winter and the length of this period increases with latitude [Brustad et al 2004]. In Tromsø, North-Norway at 70°N, cutaneous vitamin D synthesis may be absent from early October to mid-March [Engelsen et al. 2005]. People living in areas with no vitamin D effective UVR for long periods are dependent on dietary sources of vitamin D. Sun holidays and the use of sun beds have been found to be strong predictors of their 25(OH)D levels [Brustad et al. 2004].

1.3.2 Vitamin D metabolism

Vitamin D are lipophilic molecules and in the circulation most vitamin D metabolites are transported bound to the vitamin D binding protein (DBP) [Jones 2008]. Once in the circulation, vitamin D is either transported to be stored in fat tissues or to the liver to be converted to 25(OH)D by one or several cytochrome P450s [Adams and Hewison 2010]. CYP2RI appears to have the highest affinity for vitamin D [Adams and Hewison 2010], and to be the most important 25-hydroxylase involved [Dusso *et al.* 2005]. Mutations in CYP2RI genes have been identified in patients with low 25(OH)D [Dusso *et al.* 2005]. The conversion of vitamin D to 25(OH)D is hardly regulated and the levels of 25(OH)D reflect the solar and dietary exposure [Rosen 2011;Dusso *et al.* 2005]. The half-life of 25(OH)D is about 15 days [Jones 2008] and because of its stability and its reflection of vitamin D from all sources, it is the preferred metabolite for measuring nutritional vitamin D status [Ross *et al.* 2011].

25(OH)D is a prohormone to the active form of vitamin D, the hormone 1,25dihydroxyvitamin D (1,25[OH]₂D) [Adams and Hewison 2010], which is synthesised principally in the kidney by the enzyme 1α -hydroxylase [Dusso et al. 2005]. The renal 1α hydroxylase is highly regulated due to its potent activity regulating the calcium homeostasis, and the half-life of 1,25(OH)₂D is only 10–20 hours [Jones 2008]. Hypocalcaemia may either directly, but especially through stimulating PTH production, enhance the synthesis 1,25(OH)₂D [Rosen 2011;Dusso et al. 2005;Adams and Hewison 2010]. Enhanced synthesis is also promoted by low phosphate concentration. The resulting increase in 1,25(OH)₂D promotes intestinal calcium and phosphate absorption, and mobilizes calcium and phosphate from bone matrix [Adams and Hewison 2010]. When the mineral concentration is corrected, fibroblast growth factor 23 (FGF-23) secreted by osteocytes in the bone matrix downregulates the synthesis of 1,25(OH)₂D [Adams and Hewison 2010;Rosen 2011]. There is also a negative feedback regulation of PTH by 1,25(OH)₂D, minimizing the potential for vitamin D intoxication [Dusso et al. 2005]. Figure 5 illustrates the synthesis and metabolism of vitamin D. Activation of vitamin D does not only occur in the kidneys. Extrarenal hydroxylation also occurs in other tissues; bone, colon, breast, uterus, ovary, prostate, lung, pancreas, monicytes/macrophages, blood vessels and synovial cells, and parathyroid cells [Dusso et al. 2005;Peterlik and Cross 2005]. This system, where 1,25(OH)₂D locally acts as a cytokine, is not regulated by PTH, but by the supply of the substrate 25(OH)D [Adams and Hewison 2010]. The different vitamin D metabolites and especially the most potent 1,25(OH)₂D, requires a mechanism to attenuate its activity. Vitamin D is potentially toxic, and the toxicity results from hypercalcemia, nephrocalcinosis, aortic calcification and other unwanted deposits of calcium and phosphorous in soft tissue [DeLuca 2008]. When adequate amounts of vitamin D is available, both 1,25(OH)₂D and 25(OH)D are catabolised to biologically inactive, water-soluble calcitroic acid [Holick 2007] and this is carried out in virtually al target cells [Dusso et al. 2005].

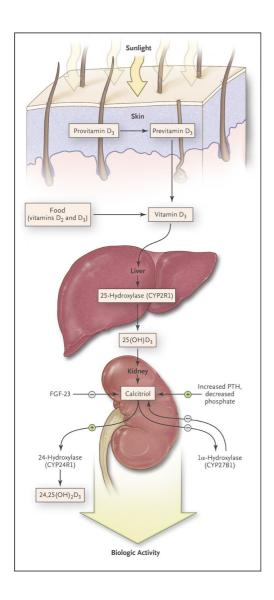


Fig. 5: Synthesis and Metabolism of Vitamin D. Reproduced with permission from [Rosen 2011], Copyright Massachusetts Medical Society

1.3.3 Vitamin D effects

Most of the biological activities of vitamin D require a VDR on the target cells, which acts as a ligand-activated transcription factor and regulates transcription of vitamin D- responsive genes [Dusso *et al.* 2005]. Such VDR are found in many genes, mainly related to bone metabolism, but 1.25(OH)₂D is also involved in controls of more than 200 genes responsible for regulation of cellular proliferation, differentiation, apoptosis and angiogenesis [Holick 2007]. The main effect is in the vitamin D endocrine system, where vitamin D acting as a hormone is an essential component maintaining and regulating the mineral homeostasis and especially keeping extracellular calcium levels within narrow limits: a process vital for normal cellular physiology and critical for skeletal mineralisation [Dusso *et al.* 2005;Rosen

2011]. 1.25(OH)₂D is also a potent immunomodulator, especially modulating the immune response to invading microbial agents [Adams and Hewison 2010]. Essentially all published studies report inhibition of Th1-associated cytokine production and a modulation of the Th2 response, affecting cytokines such as IL-4, IL-5 and IL-13 [Lange *et al.* 2009]. It has also been shown to upregulate T-regulatory cells, leading to an increase in the anti-inflammatory cytokine IL-10 [Lange *et al.* 2009]. In the past years, attention has also turned to non-skeletal effect of vitamin D, particular in relation to cardiovascular disease, cancer and immune dysfunction [Pearce and Cheetham 2010;Holick 2007].

1.3.4 Vitamin D levels and intake

Vitamin D deficiency is defined by 25(OH)D < 25 nmol/l [Meyer et al. 2006;Ross et al. 2011]. It may manifest as a medical condition with skeletal deformities known as rickets in children, and osteomalacia in adults. It causes pains in bone and muscles and muscle weakness. Affected children have difficulty standing and walking, and elderly more frequent falls and fractures [Holick et al. 2011]. The optimum level of 25(OH)D however, is more frequently debated. The US and Canadian Institute of Medicine (IOM), based on bone health outcomes, have suggested that 25(OH)D levels of at least 50 nmol/l are sufficient [Ross et al. 2011] and this is also the recommended 25(OH)D level by the Norwegian Directorate of Health [Meyer et al. 2006]. Being highly hydrophobic and tightly protein-bound, measuring serum 25(OH)D is challenging [Hollis 2008]. There are several 25(OH)D assays available, and they can mainly be divided in two groups; protein-binding assays (PBA) including competitive protein-binding (CPBA) assay and radioimmunoassay (RIA) and direct physical detection methods including high-performance liquid chromatography (HPLC) and liquid chromatography coupled with mass spectrometry (LC/MS) [Hollis 2008]. In the immunoassays, an antibody will recognize 25(OH)D, and the quantification relies on enzymatic, radioactive or electrochemilumnicent marker coupled to the antibody. The RIA have been used in the majority of the studies worldwide to define "normal" 25(OH)D levels, but both PBA may have difficulties with matrix effects and cross-reactions between the various vitamin D metabolites [Binkley et al. 2009]. The latest addition to automated 25(OH)D assay is from Roche Diagnostic, their test is an RIA and can be performed on their Elecsys and Cobas system [Hollis 2008]. Results from the Tromsø study have revealed an impact of smoking on the results of serum 25(OH)D analysis using this assay [Grimnes et al. 2010]. The HPLC separate the vitamin D₂ and Vitamin D₃ and is by most people considered the gold standard [Hollis 2008]. However, the equipment is very expensive and the throughput is much lower than for the automated immunoassays [Hollis 2008]. To define the optimal 25(OH)D levels, several markers for skeletal and extraskeletal outcome have been used. There is an inverse association between 25(OH)D and PTH, and the 25(OH)D level needed to maximally suppress the PTH concentration has been investigated in several studies [Dawson-Hughes et al. 2005]. Threshold points of 25(OH)D ranging from 25 nmol/l to 125 nmol/l have been reported, a recent three-face study in a cohort of 387 healthy Caucasian men and women aged 65 and above showed a rapid-change phase up to 25 nmol/l, a slow-change phase up to 70 nmol/l and a no change phase at 25(OH)D of 70 nmol/l and higher [Durazo-Arvizu et al. 2010]. Because of its partial dependence upon vitamin D, calcium absorption is another functional indicator of vitamin D adequacy. At 25(OH)D level of 50 nmol/l calcium absorption was found significantly reduced relative to that at 25(OH)D level of 86 nmol/l [Heaney et al. 2003]. Vitamin D, through its effect on the calcium metabolism, is linked to bone health outcomes (BMD, fracture, fall and muscle strength). Higher 25(OH)D have been associated with reduced rates of bone loss, falls and fracture [Dawson-Hughes et al. 2005] and significant positive association between 25(OH)D levels up to 90 nmol/l and BMD have been found [Bischoff-Ferrari et al. 2004]. Therefore some experts recommend serum 25(OH)D values of at least 70- 75 nmol/L for bone health [Dawson-Hughes et al. 2005; Holick et al. 2011]. For non-skeletal outcomes the available scientific evidence is found insufficient to support any recommendation of sufficient 25(OH)D level [Ross et al. 2011]. Recommended dietary intake corresponding to the recommended level of serum 25(OH)D level of 50 nmol/L is according to the IOM 600 International Units (IU) daily for ages 1-70 years, 400 IU for infants and 800 IU for ages 71 and older [Ross et al. 2011], while the Norwegian recommendation is 400 IU daily for ages 6-23 months and those older than 60 years and 300 IU for ages 2-60 years [Meyer et al. 2006]. In the report by the IOM, the limit for upper tolerable daily intake is 4000 IU and there may be reason for concern at serum levels \geq 125 nmol/L [Ross *et al.* 2011]

1.4 Multiple sclerosis, osteoporosis and vitamin D

1.4.1 Shared pathogenic and aetiological factors for MS and osteoporosis

Low circulating serum 25(OH)D levels predispose to low BMD and osteoporosis, and in severe form it may lead to vitamin D-dependent rickets [Meyer *et al.* 2006;Ross *et al.* 2011]. The hormone 1,25(OH)D₂ is, in addition to be the main regulator of the calcium homeostasis, also a potent immune regulator [Smolders *et al.* 2008a] and through this effect

hypovitaminosis is a candidate risk factor for MS [Correale et al. 2009]. Since vitamin D status may be a risk factor for MS, skeletal consequences of hypovitaminosis D could be apparent from the onset of the disease [Moen et al. 2011b]. Osteoimminology and the immunepathogenesis of MS also share several proinflammatory cytokines (IL-1, TNF-α, IL-6, IL-11) [Altintas et al. 2009]. Abnormalities in the OPG/RANKL system regulating the osteoclastogensis have been reported in various immune diseases and a small study reports significantly higher OPG and RANKL in MS patients than in healthy controls [Kurban et al. 2008]. High serum levels of the proinflammatory cytokine, osteopontin (OPN), predict the development of osteoporosis in post-menopasusal women [Chang et al. 2010] but increased levels have been reported in MS brain lesions and in plasma and CSF in MS patients [Braitch and Constantinescu 2010; Vogt et al. 2003]. Smoking is also regarded as a risk factor for both MS and osteoporosis [Compston and Coles 2008; Poole and Compston 2006]. Limited physical activity and low level of exercise due to immobility in persons with MS may interfere with the bone mass acquisition and cause a higher bone loss than in healthy individuals. Daily use of corticosterioids during three or more months is a known risk factor for osteoporosis also in MS patients [Stepan et al. 2004]. Treatment with pulsed intravenous metylprednisolon does not seem to confer the same risk [Dovio et al. 2004;Zorzon et al. 2005], although Triantafyllou found that BMD measurements was negatively associated with increasing dosage of intravenous corticosteroids [Triantafyllou et al. 2012].

1.4.2 BMD and osteoporosis in persons with MS

Due to the relationship between immobility and osteoporosis, Nieves and coworkers first proposed that MS patients may be at increased risk for osteoporosis and fracture. In a study of female MS patients they found that both total body bone mineral (TBBM), BMD in the lumbar spine and in the femoral neck were related to the severity of the MS [Nieves *et al.* 1994]. Later several studies have measured BMD in MS patients. A comprehensive review of 20 published studies and one unpublished study retrieved on 5 April 2011 was published by Gibson and Summers [Gibson and Summers 2011]. Five out of six case-control studies showed reduced BMD in people with MS in either lumbar spine, femoral neck or total body. Only one study found no reduction [Zorzon *et al.* 2005]. In studies comparing BMD in persons with MS with reference data, the overall picture is that BMD is reduced at the lumbar spine and hip. A recently published case-control study reports a statistically significant lower BMD at femoral sites compared to healthy controls [Sioka *et al.* 2011]. A consistent finding with a strong negative correlation with EDSS where found, and the correlation was strongest

at the femoral neck. Most studies report BMD measurements as mean average T- or Z-scores, but some studies report the prevalence of osteopenia and osteoporosis. One study of men with MS found osteopenia in 43% and osteoporosis in 38 % of the participants [Weinstock-Guttman *et al.* 2004]. In a group of 70 persons with MS on long-term treatment with glucocorticoids, 46% had osteopenia and 45% osteoporosis [Stepan *et al.* 2004]. In a small case-control study, respectively 9.6% and 38.4% of ambulatory premenopausal women with MS had osteoporotic and osteopenic BMD values, in the control group the respective values were 0% and 26.8% [Terzi *et al.* 2010]. The latest study of 119 RRMS patients, osteopenia in lumbar spine and femoral neck was present in 26% and 50%, and the number for osteoporosis was 3% and 11% respectively [Triantafyllou *et al.* 2012].

A likely explanation for the association with EDSS is the immobility and loss of mechanical loading which may accelerated bone loss [Raisz 2005;Poole and Compston 2006], however a recently published case-control study in people with CIS or MS shortly after MS onset, also found that 50.5% of the persons with CIS or MS shortly after onset had either osteopenia or osteoporosis, compared to 37.1% of the controls, even though the persons with MS had no or minor physical disability [Moen *et al.* 2011b].

1.4.3 Fracture risk in persons with MS

The incidence of fractures and the risk factors for fractures during the course of MS have been described in several studies, and the results presented until April 2011 have been reviewed by Gibson and Summer [Gibson and Summers 2011]. They describe four observational studies with several limitations where the annual fracture risk was between 1.3% and 6.2 % [Weinstock-Guttman et al. 2004;Troiano et al. 1992;Logan, Jr. et al. 2008;Sibley et al. 1991]. Only one of the studies which included 170 persons with MS reported an increased annual fracture incidence compared to the general population annual fracture incidence, which has been estimated at 3.6% by Donaldson and coworkers [Donaldson et al. 2008]. Case-control studies, however, suggest an increase in annual fracture incidence in persons with MS, and cross sectional data have shown an increase in fracture rate with increasing levels of disability [Gibson and Summers 2011]. A population-based case control study where one aim was to compare the history of fracture in people with CIS or MS shortly after MS onset compared to the general population found, in spite of enhanced self-reported fall tendency, no difference in the number of fractures reported [Moen et al. 2011a]. In a population-based cohort study in the United Kingdom, using data from the UK General Practice Research Database linked to the National Hospital Registry, they found that compared to controls, MS patients had an

almost three fold increased risk of hip fracture and a 1.4 fold increase in osteoporotic fracture in general [Bazelier *et al.* 2011]. In a case control study where the case population was retrieved from the Danish MS Registry linked to the National Hospital Discharge Register, incidence rate ratio (IRR) between MS patients and controls was 1.4, where in particular the risk of tibia fracture (IRR 3.36), femur fracture (IRR 6.66) and hip fracture (IRR 3.20) were elevated [Bazelier *et al.* 2012b]. From the same Danish register incident MS persons with MS were indentified, and the study showed that incident MS patients had an increased risk of femur/hip fracture, a risk that was almost doubled compared to controls [Bazelier *et al.* 2012a].

The increased risk of fracture in persons with MS may be caused by the underlying conditions such as hypovitaminosis D, limited physical activity, use of corticosteroids and other medication, and smoking (se section 1.4.1). In addition it is generally accepted that MS increases the risk of falling due to the symptoms of the disease including imbalance, muscle weakness, blurred vision, dizziness and cognitive impairment [Bazelier *et al.* 2012c;Moen *et al.* 2011a]. In the review by Gibson and Summers fall frequency ranged from 32% reporting ≥ two falls in the past 2 month to 64% reporting ≥ two falls in 1 year, which are high compared to fall frequency in elderly of whom approximately 30% fall annually [Gibson and Summers 2011]. Moen et al found that 20% of the persons with CIS or recently diagnosed with MS actually had reported a tendency to fall, against only 3% in controls. Propensity to fall was associated with EDSS [Moen *et al.* 2011a].

2. Aims of the study

The aim of this phase II 96 week randomized clinical trial (RCT) was to assess whether a weekly dose of 20,000 IU vitamin D_3 could prevent bone loss in fully ambulatory persons with MS age 18–50 years.

The primary objective of the study was to determine changes in BMD over the study period comparing the treatment group and the placebo group.

The secondary objectives were to assess:

- The prevalence of low BMD at baseline (compared with standard laboratory reference values)
- Which past and current exposures that were associated with baseline BMD

- The vitamin D status at baseline and its predictors
- The effect of supplementation of 20,000 IU Vitamin D₃ per week on vitamin D status
- The seasonal variations of 25(OH) vitamin D in the placebo group

3. Methods and materials

3.1 Trial design

3.1.1 Participants

This was a single centre, randomised, double-blinded, placebo-controlled, 96 week trial designed to assess the effect of supplementing 20,000 IU vitamin D_3 weekly on bone mineral density in fully ambulatory patients with relapsing-remitting multiple sclerosis. The study was conducted at the Department of Neurology, University Hospital of North Norway, the only neurology service in Troms and Finnmark, the northernmost counties of Norway. The department manages approximately 200 persons aged 18-50 years with a diagnosis of MS according to the McDonald criteria [McDonald *et al.* 2001] in a population of 250,000 people from Troms, Finnmark, and northern Nordland above the Arctic Circle. From this population, 153 patients who were assumed to be able to walk without rest for 300 m, were invited to participate. Patients were eligible for inclusion if they were 18 to 50 years of age with clinical definite MS according to McDonald criteria and EDSS score \leq 4.5 [Kurtzke1983].

3.1.2 Exclusions criteria

Exclusion criteria were in paper I: Not able to walk 300 m; postmenopause; other autoimmune illness; history of conditions or diseases affecting bone; pregnant or lactating during the past 6 months; use of bone-active medications

Exclusion criteria were in paper II and III: As in paper I except inability to walk 500 m or more and unwillingness to use appropriate contraception. A pregnancy test (HCG in serum) was performed in all female participants at screening.

3.1.3 Ethics

The study was approved by the Regional Committee for Medical and Health Research Ethics. All participants provided written informed consent. The trial was registered in ClinicalTrials.gov (ID NCT00785473).

3.2 Randomisation and intervention

A statistician at the Clinical Research Centre, who was not otherwise involved in the study, performed the randomisation by blocks of six, stratified by sex, with a concealed, computergenerated randomisation procedure. An identification number and a randomisation number were created for each participant. All study personnel and participants were blinded to treatment assignment for the duration of the study. The active treatment was 20,000 IU vitamin D₃ (cholecalciferol) once a week, administered as a capsule of DekristolTM (Mibe GmbH Arzneimittel, Brehna, Germany). The average cholecalciferol content measured in 10 capsules was 18,120 IU per capsule, which is within the 10% limit accepted by EU authorities. Identical placebo capsules were provided by the manufacturer of DekristolTM (SWISS CAPS AG, Kirchberg, Switzerland). Participants were allowed to continue vitamin D supplements they used at baseline. All participants received 500 mg elemental calcium daily, administered as a chewable tablet of Weifa KalsiumTM (calcium carbonate, Weifa AS, Oslo, Norway). Participants who had gastrointestinal side effects attributed to Weifa calcium discontinued the calcium supplement if they had an estimated dietary calcium intake of ≥ 800 mg/day. If calcium intake was lower, they switched to Calcium Sandoz™ effervescent tablets (calcium lactate-gluconate and calcium carbonate, Sandoz A/S, Odense, Denmark) which were better tolerated.

3.3 Adherence and safety

The participants were offered a reminder to take the study medication by weekly mobile phone text messages. Adherence to study medication was evaluated by capsule counting (Adherence [%] = [number of capsules consumed/number of capsules that should have been consumed] x 100). The participants were considered adherent if they had taken at least 80 % of the study medication. Participants were educated regarding clinical signs of hypercalcaemia, and ionised serum calcium was determined every 12 weeks. Adverse events were assessed every 12 weeks via phone consultation, at study visits week 48 and week 96, and in the event of hospital admission. Serious adverse events and unexpected medical events were reported to the Norwegian Medicines Agency and to the Regional Committee for Medical and Health Research Ethics.

3.4 Clinical examination

Full physical examination was performed and complete neurological status from which EDSS was determined at each study visit by a neurologist. Erect height and body weight were

measured with participants wearing light clothing and no shoes. The MS functional composite (MSFC) was assessed by trained nurses [MSFC administration and scoring manual, 2001]. Strength of handgrip (kPa) was determined using a Martin vigorimeter (Elmed Inc., Addison, IL, USA). Variables that were measured at study visits or retrieved from charts are listed in table 1.

3.5 BMD measurements

Measurement of BMD by DXA was performed by trained technicians using a Lunar Prodigy advanced densitometer (Lunar Radiation Corp., Madison, WI, U.S.A.). The long-term precision was 0.26-0.28%, obtained by daily calibration of the densitometer. BMD was determined at the hip (mean of left and right total hip), the spine (anterior-posterior spine L1-L4), and the non-dominant ultradistal (UD) radius. In clinical practice, BMD is commonly measured at the hip and the lumbar spine. We added BMD at the forearm in non-dominant hand, which also is a relevant fracture site, to assess possible effects of MS on a peripheral skeletal site.

Table 1 Variables measured at study visit, retrieved from charts, or collected by questionnaire

Demographic and anthropometric	Physical function and activity	MS-related	Childhood & adolescence (retrospective)	Nutritients, medications, hormones, lifestyle habits
Age Sex Years lived above the Arctic Circle before age 20 Family history (1 st degree relatives) of low energy fractures* (yes/no) Personal history of low energy fractures* (yes/no) Body mass index (BMI, kg/m²)	Ambulatory function without rest or aid (>500 m/≤500 m) Timed 25-foot walk (sec)[2001] 10-foot timed tandem walk (sec)[Herbert 2006] Strength of grip nondominant hand (kPa) 9-hole peg test nondominant hand (sec)[MSFC administration and scoring manual, 2001] Leisure time physical activity [†]	Age at MS onset Disease duration EDSS [Kurtzke1983]	Physical activity [†] Consumption of milk and yoghurt (glasses or cups/d) Cod-liver oil supplement use (no/winter only/all year) Summer outdoor activities age 16-20 [‡] Body shape at age 15 [§] Eating disorder (yes/no)	Serum 25(OH)D (nmol/L) Vitamin D intake from diet and supplements (µg/d)¶ Solarium past year (yes/no) Calcium intake from diet and supplements (mg/d)¶ Cumulative dose of intravenous methylprednisolone (g) Ongoing beta-interferon treatment (yes/no) Age at first menstruation Oral contraceptive use (years) Alcohol (U/week) Smoking (years daily smoking)

*Fracture resulting from low energy trauma (fall from standing height or less, or trauma that in a healthy individual would not give rise to fracture); † Physical activity was categorised as: (a) mostly sedentary; (b) walking or bicycling at least 4 h/week; (c) exercising at least 4 h/week; and (d) participating in competitive sports several times a week; ‡ In summer, how much did your activities (playing, sports, walking, working) take you outside? (a) not that often, (b) a moderate amount, (c) quite a lot, (d) virtually all the time [van der Mei *et al.* 2007]; § rated on nine-level figure drawing [Michels *et al.* 2007]; § assessed by food frequency questionnaire [Hjartaker *et al.* 2007].

3.6 Blood tests

Every 12 weeks blood was drawn and haematology, liver enzymes, ionised calcium and PTH were analysed at the Department of Clinical Chemistry, University Hospital of Northern Norway. For interpretation of these blood tests, laboratory reference ranges were used. At week 96 serum testosterone was measured in men and FSH in women. FSH values of ≥ 40 IU/L were defined as a marker for perimenopause [Harlow et al. 2007; Recker et al. 2000]. Batch analyses of baseline serum 25(OH)D were first performed using an electrochemiluminescence immunoassay on a CobasTM analyser (Elecsys vitamin D₃ reagent kit, Roche Diagnostics GmbH, Mannheim, Germany), and these results are presented in paper I. Analyses of serum 25(OH)D measurements presented in paper II and III were performed by mass spectroscopy (coefficient of variation at 75 nmol/L was 3,5%) at the Hormone Laboratory, Haukeland University Hospital, Bergen, Norway. In the placebo group, analyses were performed at baseline (winter), at study week 60 (spring), 72 (summer), 84 (autumn) and at study end (winter). In the intervention group, serum 25(OH)D levels were only measured in winter. Serum 25(OH)D levels were classified as deficient (<25 nmol/L), insufficient (25-50 nmol/L), sufficient (\geq 50 nmol/L), and high (\geq 75 nmol/L). We applied the level of sufficiency as recommended by the IOM and the Norwegian Directorate of Health (50nmol/L) in statistical analyses [Ross et al. 2011; Meyer et al. 2006].

3.7 Questionnaires

The information collected by self-administered questionnaires is specified in Table I. Dietary vitamin D intake was calculated from a validated food frequency questionnaire, administrated at week 0, 48 and 96 [Brustad *et al.* 2004;Hjartaker *et al.* 2007]. Use of vitamin D-containing supplements and sun exposure during the last three months (tanning bed and sun vacation) were recorded every 12 weeks. Time spent outside during the last 7 days was reported in spring, summer and autumn, but not in winter. At 69° N the sun is below the horizon from November 20th to January 20th, and ambient UVR is not sufficient for cutaneous vitamin D synthesis from early October through mid-March [Engelsen *et al.* 2005]. Physical activity was categorised as: a) mostly sedentary, b) walking/bicycling at least 4 h/week, c) exercising at least 4 h/week or d) participating in competitive sports several times a week.

3.8 Statistical analysis

Data from BMD measurements in a comparable group of persons with MS were not available when the trial was planned, and power calculation was not performed prior to the study.

Analyses were carried out according to a pre-established analysis plan. In the patient who discontinued the study medication because of pregnancy, the last measurements (week 48) were carried forward. Comparisons between groups were made using Student's T test for continuous data, Mann-Whitney U test for data that were not normally distributed and χ^2 test/Fisher exact test for categorical variables. Pearson correlation coefficients (r) were calculated for bivariate correlations. In paper II and III the modified intention-to-treat population included all participants who received at least one dose of the study medication and presented at one follow-up visit. All statistical analyses were conducted using SPSS 15.0, 16.0 and 18.0 for Windows (SPSS Inc. Chicago, IL, U.S.A.). P-values < 0.05 were considered statistically significant.

3.8.1 Statistical tests applied in paper I

Multiple linear regression models were constructed for the dependent variables "BMD femoral neck", "BMD lumbar spine", and "BMD non-dominant UD radius". Variables introduced into regression models were those with P values < 0.25 in Pearson correlations. Five initial models were built for each site of measurement. Integrated models for each site included sex, BMI and variables that were significantly associated with BMD at the particular site of measurement in the initial models. The final models were developed considering all variables that showed significant associations in at least one of the integrated models. Logistic regression was applied to the dependent variable "low BMD", according to the same principles of modelling as described for linear regression. The high number of variables available for regression analysis in relation to the number of participants in the study carries a risk of finding spurious associations. Formal corrections for multiple measurements were not applied, but results were interpreted with caution when P values were between 0.05 and >0.005. Where judgments of statistical significance were made, the biological feasibility and consistency of the findings were taken into account.

3.8.2 Statistical tests applied in paper II

Distribution and variance of BMD and vitamin D values were assessed by appropriate tests. Interactions between sex and group were tested for. The main outcome, percentage of change in BMD, was analysed by independent sample T-test. Changes in BMD from baseline to week 96 were analysed by paired samples T-test. A preplanned multiple regression model included the variables age, body mass index (BMI), baseline BMD, baseline 25(OH)D, percentage

change in 25(OH)D, and perimenopausal FSH values. Subgroup analyses were performed by sex and by treatment group.

3.8.3 Statistical tests applied in paper III

Comparisons of baseline characteristics were performed between treatment groups, between dichotomised 25(OH)D levels (< or \ge 50 nmol/L at baseline), and between women and men. Univariate analyses of associations were performed for baseline 25(OH)D values and baseline characteristics. Predictors of 25(OH)D on population level, age, BMI and sex, as well as possible predictors of 25(OH)D values (p value <0.25 in univariate analysis) were included in linear regression models. The paired Student's T-test was used to compare mean 25(OH)D values at baseline and at study end in both treatments groups, and between winter and summer values in the placebo group.

4. Summary of the results

4.1 Paper I and paper I erratum

In this paper we present the predictors and the prevalence of low bone mineral density in fully ambulatory persons with multiple sclerosis. 80 participants were included in the analysis. 24% (20% women and 34% men) had low BMD (z-score ≤ -2.0) in one or several measured sites. Mean BMD for women and men at femoral neck, lumbar spine and UD radius are shown in figure 6. BMD at femoral neck was significantly negatively associated with age, male sex, slower 10-foot timed tandem walk and positively associated with BMI, walking distance > 500 m and physical activity growing up. In female participants there was also a positive association with oral contraceptive use. According to BMD at the lumbar spine we found the same associations, except for age, slower 10-foot timed tandem walk and use of oral contraceptives. BMD at the non-dominant UD radius was positively associated with BMI, strenght of grip, age at MS onset and summer outdoors activity at age 16-20 year. Serum 25(OH)D levels were not correlated with BMD at any site.

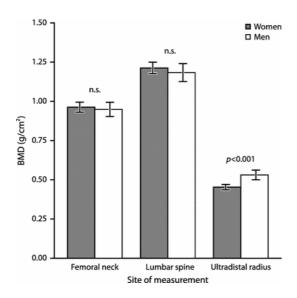


Fig. 6: Bar chart showing BMD for women and men at three sites of measurements. Error bars represent 95% confidence intervals.

4.2 Paper II

In this paper we report the results with regard to the main objective and the aim of this study whether a weekly dose of 20,000 IU vitamin D₃ could prevent bone loss in fully ambulatory persons with MS age 18–50 years. Figure 7 shows the flow of the participants in the study. Percentage change in BMD from week 0 to week 96 did not differ between participants who had received vitamin D₃ supplementation and those who had received placebo. This result was not altered by adjustment for baseline serum 25(OH)D or change in serum 25(OH)D over the study period. There were no significant associations between percentage change in BMD and possible predictors of BMD (sex, age, BMI, baseline BMD, baseline serum 25[OH]D, change serum 25[OH]D, serum testosterone in men at week 96, and EDSS). One participant in each treatment arm experienced an unexpected medical event that was not related to the study medication and one participant in the placebo group discontinued the study medication at week 94 due to nephrolithiasis and ionised calcium (iCa) during the study did not exceed the upper limit of reference values (1.34 mmol/L) in any of the participants.

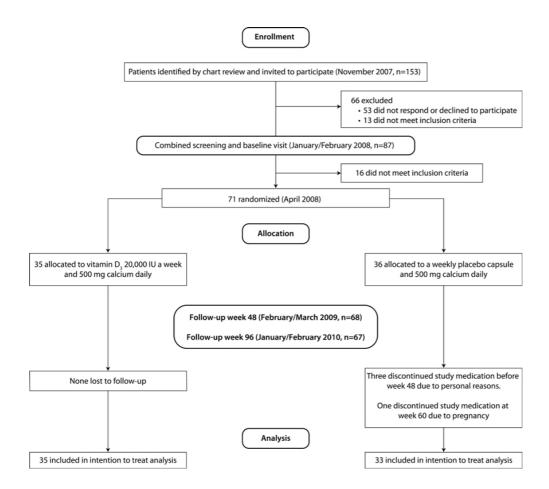


Fig. 7: Flow of the participants through the study

4.3 Paper III

In this paper we report vitamin D status and its predictors in persons with MS living at high northern latitude (68°-71°). At baseline during winter 10% of the patients were vitamin D deficient (serum 25[OH]D <25 nmol/L), 40% were deficient (serum 25[OH]D between 25 and 49 nmol/lL), 29 had sufficient levels (serum 25[OH]D between 50 and 74 nmol/lL) and 21% had high levels (serum 25[OH]D >74 nmol/L). Mean serum 25(OH)D was 56 nmol/L and was positively predicted by sun vacation last three months, tanning bed last three months and total vitamin D₃ intake. Mean total daily intake of vitamin D₃ for the participants with serum 25(OH)D< 50 nmol/l was 6.5 μ g (95% CI 5.2;7.8) (260 IU [95% CI 208;312]) and for the participants with serum 25(OH)D \geq 50 nmol/l 13.3 μ g (95% CI 9.8;16.9) (532 IU [95% CI 392;676].

In the placebo group we found a seasonal variations in serum 25(OH)D levels with a mean 25(OH)D level during baseline winter at 58 nmol/L that raised to 87 nmol/L during summer, and the proportion of the participants with sufficient 25(OH)D levels (≥ 50 nmol/L) increased

from 55% in winter to 92% during summer. In the intervention group mean serum 25(OH)D increased from 56 nmol/L at baseline to 123 nmol/L at study end. This represents a mean increase of 2.3 nmol/L per 100 IU vitamin D_3 daily supplement, and all participants achieved sufficient vitamin D levels (serum 25[OH]D > 50 nmol/L) at study end and 91% had serum 25(OH)D values \geq 75 nmol/L. Figure 8 shows the seasonal variation in serum 25(OH)D in the placebo group and at study start and study end in the intervention group.

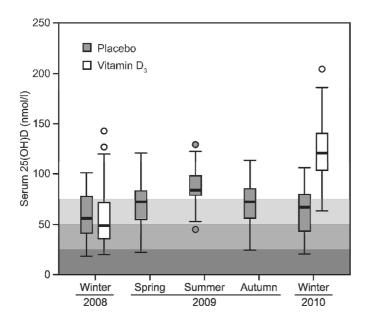


Figure 8 Seasonal variation of serum 25(OH) D levels in the placebo group and serum 25(OH)D levels at baseline and at study end in the intervention group.

5. Discussion

5.1 General discussion

This study shows that low BMD was prevalent at baseline, and a weekly dose of 20,000 IU vitamin D_3 administrated for 96 weeks could not prevent bone loss in fully ambulatory persons with MS age 18-50 year. Findings did not vary by sex and were not altered after adjustment for serum 25(OH)D. MS-related variables, as well as lifestyle exposure during growing up, differentially affect BMD at three clinically important sites: the femoral neck, the lumbar spine and the UD radius. At the study start in winter, mean serum 25(OH)D was 56 nmol/l, and half of the participants had sufficient 25(OH)D levels (\geq 50 nmol/l). Winter serum 25(OH)D levels were strongly predicted by sun exposure through sun vacation or solarium and by vitamin D intake. In the placebo group, the proportion of study participants with

sufficient levels of vitamin D increased from 55% during winter to 92% during summer. All participants in the intervention group achieved sufficient (\geq 50 nmol/l) winter levels of serum 25(OH)D.

5.1.1 Paper I

At the combined screening and baseline visit, low BMD (z score \leq -2) was present in 19 out of 80 participants. Because BMD follows a Gaussian distribution, low BMD (z score \leq -2.0) would be expected to be present in approximately 2.5% of premenopausal women and men below age 50 [Lewiecki 2005]. Prevalence of low BMD was much higher in our study, where almost 25% of the participants at screening had low BMD at one or more sites. Previous studies reporting osteopenia (t-score between -1.0 and -2.5) and osteoporosis (t-score \geq -2.5) present even higher number [Altintas *et al.* 2009;Stepan *et al.* 2004;Weinstock-Guttman *et al.* 2004;Moen *et al.* 2011b;Triantafyllou *et al.* 2012]. We further demonstrate that MS-related variables as well as past exposures differentially affect BMD at three clinically important skeletal sites: the femoral neck, the lumbar spine, and the UD radius. BMI is a strong and well known predictor of BMD, and we found that BMI was positively associated with BMD at femoral neck, lumbar spine and UD-radius. Most published studies have reported BMD without adjustment for BMI, but in a recently published study by Sioka et al, BMI was a significant independent variable of BMD values in lumbar spine and femoral neck in persons with MS [Sioka *et al.* 2011].

BMD at the femoral neck and lumbar spine were associated with current ambulatory function and physical activity during growth, indicating a major effect of exercise and mechanical loading at these skeletal sites. In a review by Gibson and Summers a consistent finding was a strong negative correlation between EDSS and BMD level, where the correlation was strongest at the femoral neck [Gibson and Summers 2011]. However, we found that ambulatory function (>500 m/≤500 m) resulted in a better fitting model than EDSS at the femoral neck and lumbar spine. We also found that the 10-foot timed tandem walk was independently associated with BMD at the femoral neck. This test reflects mild ambulatory impairment, related to impaired balance, better than walking distance. Persons with impaired balance may engage in less physical activities with high mechanical loading that are crucial to maintain BMD. BMD was not associated with the present level of physical activity. This is in agreement with a population-based study showing that physical activity during growth, but not current physical activity influenced BMD [Bainbridge *et al.* 2004]. However, high-impact exercise has been shown to significantly increase BMD at the hip in a randomized controlled

trial [Vainionpaa *et al.* 2005]. Age was negatively associated with BMD only at the femoral neck, the site at which bone loss starts first [Vondracek *et al.* 2009].

BMD at the UD radius, a clinically relevant site, has not previously been explored in persons with MS. We found a strong association with strength of grip at this site, an association that also has been demonstrated in healthy individuals [Aydin *et al.* 2006]. Arm dominance has a pronounced effect on bone density, especially if the individual is involved in any type of repetitive unilateral activity. Therefore measurements from the non-dominant arm are used in studies [Bonnick 2004]. Summer outdoor activities age 16–20 were associated with BMD at the UD radius. Interestingly, this measure, a proxy of vitamin production in the skin [van der Mei *et al.* 2006], was associated with MS risk in the same population [Kampman *et al.* 2007]. The difference in variables predicating BMD in UD radius compared to those at the femoral neck and lumbar spine may be a consequence of the UD radius not being weight-bearing, which might result in increased susceptibility to other influences.

Peak bone mass is higher in men than in women [Poole and Compston 2006]. To our surprise, male rather than female sex showed a possible negative association with BMD at the femoral neck and lumbar spine. In addition, at the UD radius, BMD values were only 18% higher in men than in women, compared with the 28% difference reported in a population-based study in the same geographical area [Emaus *et al.* 2005]. Previous studies reporting BMD values for both men and women with MS found results similar to ours, but these findings were not commented on by the authors [Cosman *et al.* 1998;Stepan *et al.* 2004;Tuzun *et al.* 2003]. A recent published case control study report that in men with MS the mean BMD at both lumbar spine and femoral sites were lower by about 8 % than in the healthy controls [Sioka *et al.* 2011].

A causality is established between serum 25(OH)D and skeletal outcomes, including BMD [Ross *et al.* 2011], but in our study population BMD was not associated with serum 25(OH)D. However none of the participants were vitamin D deficient (25(OH)D <25 nmol/L). A correlation of vitamin D status with low BMD in persons with MS was found in patients who had lower serum 25(OH)D levels and vitamin D intake and higher EDSS scores and age than ours [Nieves *et al.* 1994]. Other studies did not find significant associations of vitamin D status with BMD in persons with MS [Cosman *et al.* 1998;Ozgocmen *et al.* 2005;Weinstock-Guttman *et al.* 2004;Triantafyllou *et al.* 2012]. Published studies of persons with MS probably lack the statistical power to confirm the association between serum 25(OH)D and BMD that has been described in healthy individuals [Ross *et al.* 2011;Dawson-Hughes *et al.* 2005].

Treatment with pulsed intravenous methylprednisolone did not affect BMD, which is in agreement with most [Nieves et al. 1994;Schwid et al. 1996;Tuzun et al. 2003;Weinstock-Guttman et al. 2004;Zorzon et al. 2005;Moen et al. 2011b;Sioka et al. 2011], but not all [Formica et al. 1997;Ozgocmen et al. 2005;Triantafyllou et al. 2012] previous studies. Interferon-beta treatment induces changes in multiple proteins and mRNAs related to bone homeostasis in MS patients [Weinstock-Guttman et al. 2006]. We did not find any association between BMD and Interferon treatment, which is supported by the results from two other recently published studies [Triantafyllou et al. 2012;Moen et al. 2011b]. Cigarette smoking and alcohol consumption are known risk factors for low BMD [Raisz 2005;Poole and Compston 2006], but in this small study we could not find such association and neither did to other studies [Moen et al. 2011b;Sioka et al. 2011]. Personal and family fracture history were neither not associated with BMD.

5.1.2 Paper II

Paper II describes the effect of supplementing 20,000 IU vitamin D₃ or placebo on the BMD measurements. There was no difference in change in BMD comparing the to treatment groups. In paper II we presented the change in the BMD measurements between baseline and study end, separately in each group. We found a significant bone loss in the hip in the placebo group and not in the intervention group, and a significant bone increase in the UD radius in the intervention group and not in the placebo group. Such within group analysis is not recommended in RCTs. However in the whole study population mean BMD in the hip decreased significantly by 1% (Paired samples T-test, P<0.01). In healthy individuals, studies have reported no significant bone loss at the hip in women before the perimenopausal years [Berger et al. 2010; Berger et al. 2008; Chapurlat et al. 2000; Henry et al. 2010; Neer 2010; Recker et al. 2000], while men may or may not have a small bone loss before age 50 [Berger et al. 2010; Emaus et al. 2009; Henry et al. 2010]. The general bone loss at the hip, a weight bearing site, may result from decreased physical activity even in fully ambulatory persons with MS [Pugliatti et al. 2008b], due to fatigue, visual problems, and impaired balance. We observed no change in BMD at the lumbar spine. This finding is in agreement with two recent studies reporting unchanged BMD at the lumbar spine over a period of respectively 96 weeks and 39 months in interferon beta-treated persons with MS [Ravnborg et al. 2010; Sorensen et al. 2009]. At the UD radius the mean BMD increased significantly by 1.8% (Paired Samples T-test, P=0.01). In healthy individuals, BMD at this site has been

reported to be stable or increasing up to age 40 in both sexes [Chapurlat *et al.* 2000;Emaus *et al.* 2009;Henry *et al.* 2010]. BMD at the UD radius, especially in the non-dominant arm, is not dependent on weight bearing activity and therefore is likely to be less affected by mild to moderate MS-related disability.

There is good evidence that vitamin D supplementation improves bone health [Bischoff-Ferrari *et al.* 2009; Cranney *et al.* 2008]. A meta-analysis concluded that the effect of vitamin D supplementation on bone health was greater in persons with low (<25 nmol/l) serum 25(OH)D than those whose serum level was normal [Tang *et al.* 2007]. Only three individuals in the intervention group of this study were overtly vitamin D deficient at baseline (25(OH)D <25 nmol/L), and participants were allowed to continue vitamin D supplements they used at baseline, both of which could have contributed to the negative findings in this trial. In women, baseline BMD at the hip was significantly higher in the intervention than in the placebo group, which may also have influenced the result of the study.

5.1.3 Paper III

In paper III we report vitamin D status and its predictors in persons with MS living at high northern latitude and the effect of weekly supplementation with 20,000 IU vitamin D₃ for 96 weeks on 25(OH)D levels. At study start mean serum 25(OH)D was 56 nmol/L and serum 25(OH)D levels were strongly predicted by sun exposure through sun vacation or tanning bed and by vitamin D intake. Half of the participants had insufficient 25(OH)D levels (<50 nmol/L) during winter, even though their mean total daily intake was at 260 IU (95% CI 208;312) which is close to the recommend daily dose (300 IU) by the Norwegian Directorate of Health [Meyer et al. 2006]. In the placebo group, the proportion of study participants with sufficient levels of vitamin D increased from 55% during winter to 92% during summer. All participants in the intervention group achieved sufficient winter levels of serum 25(OH)D. During the winter months with insufficient solar UVR to induce cutaneous vitamin D production [Engelsen et al. 2005], mean serum 25(OH)D was 56 nmol/L. This is comparable with measurements performed in fair skinned persons with MS during winter at lower latitudes, with reported values in the range of 40 to 80 nmol/L [Amezcua et al. 2012;Barnes et al. 2007; Burton et al. 2010; Cosman et al. 1998; Hiremath et al. 2009; Holmoy et al. 2009;Kimball et al. 2007;Knippenberg et al. 2011;Loken-Amsrud et al. 2012;Lonergan et al. 2011; Lucas et al. 2011; Nieves et al. 1994; Orton et al. 2008; Ozgocmen et al. 2005; Pierrot-Deseilligny 2009; Runia et al. 2012; Simpson et al. 2010; Smolders et al. 2008b; Soilu-Hanninen et al. 2005; Soilu-Hanninen et al. 2008; Stein et al. 2011; van der Mei et al.

2007; Weinstock-Guttman *et al.* 2011; Yildiz *et al.* 2011; Triantafyllou *et al.* 2012; Baarnhielm *et al.* 2012]. A meta-analysis of studies in healthy subjects all over the world during all seasons reported mean serum 25(OH)D levels of 54 nmol/L [Hagenau *et al.* 2009]. Most observational studies comparing 25(OH)D levels in MS patients and controls report on less than 50 individuals in each group and results are inconsistent [Barnes *et al.* 2007; Correale *et al.* 2009; Hiremath *et al.* 2009; Holmoy *et al.* 2009; Ozgocmen *et al.* 2005; Soilu-Hanninen *et al.* 2008; van der Mei *et al.* 2007; Triantafyllou *et al.* 2012]. The largest study to date comparing 25(OH)D levels in 1013 Swedish cases of MS and 1194 matched controls found mean values of 62.9 nmol/L in cases and 66.3 nmol/L in controls, a difference that is unlikely to be of clinical significance [Baarnhielm *et al.* 2012].

During winter, the time of the year with no natural sun exposure at 69°N, we observed that 25(OH)D levels were predicted by UVR exposure through sun vacation or tanning bed use as well as by vitamin D intake. Sufficient vitamin D levels (≥50 nmol/L) were measured in 74% of the participants who at baseline either had had sun exposure during the last three months or who were taking a vitamin D supplement. Only 25% of those who neither had had sun exposure nor were taking a vitamin D supplement were vitamin D sufficient. An association between winter 25(OH)D levels and sun vacation or tanning bed use has previously been reported in Northern Norway [Brustad et al. 2004], and an association between serum 25(OH)D levels and sunlight exposure has been described in persons with MS [Simpson et al. 2010; van der Mei et al. 2007]. Most humans are not dependent upon dietary vitamin D supplies for vitamin D sufficiency [Hollis 2005]. In our study, only 38% of the participants not supplementing vitamin D were vitamin D sufficient during winter, while 69% of those who used supplements containing at least 100 IU vitamin D3 had 25(OH)D values in the sufficient range. This finding is supported by a study in persons with MS from Ireland [Lonergan et al. 2011], but not by a study from Australia [Simpson et al. 2010; van der Mei et al. 2007]. However, supplementation with vitamin D₂ (rather than vitamin D₃) was common in Australia at the time [van der Mei et al. 2007], and the contribution of UVR to vitamin D status can be assumed to be larger than in Norway and Ireland. We did not find inverse relationship between serum 25(OH)D level and BMI as has been reported in larger studies [Rosen 2011].

In the placebo group, we analysed seasonal variation in measurements of serum 25(OH)D levels. The mean 25(OH)D level was significantly higher during the summer (87 nmol/L) than during the winter (58 nmol/L), and the proportion of study participants with sufficient levels of vitamin D (\geq 50 nmol/L) increased from 55% during winter to 92% during summer.

Similar seasonal variations have been reported in other studies of MS patients with minimal disability [Soilu-Hanninen *et al.* 2008;van der Mei *et al.* 2007]. In contrast, little or no seasonal fluctuations were observed in more disabled patients [van der Mei *et al.* 2007]. Immobility limits outdoor activities and heat intolerant patients avoid direct sun exposure, both of which reduce the contribution of cutaneous vitamin D production to vitamin D status. In the intervention group, supplementation with 20,000 IU vitamin D₃ per week for 96 weeks raised serum 25(OH)D by 68 nmol/L to 123 nmol/L. This is an increase of 2.4 nmol/L per 100 IU vitamin D₃ daily supplement, which is close to the increase of 2.5 nmol/L per 100 IU vitamin D₃ that has been described earlier [Rosen 2011]. All participants achieved the recommended 25(OH)D levels (≥50 nmol/L) for healthy subjects [Ross *et al.* 2011]. Serum 25(OH)D values of at least 75 nmol/L, as recommended for good bone health [Dawson-Hughes *et al.* 2005], were achieved by 91 % of the participants. Vitamin D₃ supplementation, at a dosage 30% below the tolerable upper intake levels of 28.000 IU per week [Ross *et al.* 2011], was well-tolerated with no clinical or biochemical side effects.

5.2 Methodological aspects of the study

5.2.1 Bias and confounding

Bias is a systematic error, which might result in incorrect estimates. It tends to influence on the results and move the results away from the "truth" and it reduces the validity of the results. In the results of the RCT, selection bias was avoided by having a representative source population and a concealed randomization. Performance bias (systematic difference of care other than the intervention of interest) and detection bias (Systematic difference in outcome assessment) was avoided by the blinding of all personnel in the study. Compliance and attrition bias is systematic difference in compliance and in withdrawals between the groups. In this the study the adherence to medication was > 80% and the withdrawal was only 3 %. To avoid the possible effect of this on the results we analyzed by modified intention to treat. Recall bias is a possible limitation of our study, in particular to the results presented in paper I and III. Assessing vitamin D intake from diet and supplementation and registration of outdoors activity growing up were performed by validated questionnaires [Hjartaker et al. 2007; van der Mei et al. 2006]. The validation of variables regarding physical activity growing up and at present, sun bed use and sun vacation last three month, personal and family history of fracture and smoking, may be limited by recall bias. Confounding is a hidden variable influencing both the outcome and the exposure, resulting in an observation of a non-causual

association. It is more likely to occur in observational than in experimental studies. In paper II, analyzing the RCT, confounding was not regarded as a problem. In paper I, and partly paper II, however, we report the results of a cross-sectional study at screening, which only gives a moment of time picture and there may be confounding variables which implies that causal relations from association cannot be drawn. To limit the possibility of confounding we performed multivariate regression analysis.

5.5.2 Random errors

A type I error means reporting a difference which is false. In paper I and III the high number of variables available for analysis in relation to number of participants carried a risk of finding spurious association. Since all analyses were on secondary outcomes, formal correction for multiple measurements was not applied, but results were interpreted with caution when pvalues were between 0.05 and 0.005. Where judgments of statistical significance were made, the biological feasibility and consistency of the findings were taken into account. In paper II a type I error affecting the primary outcome of our study would mean that we falsely claimed an effect of vitamin D supplementation on BMD even there was no treatment effect. To avoid this, analyses were performed in a preplanned multiple regression model and a strict statistical criterion was predefined to assess when a finding should be regarded significant and in that paper we set the significance level at 0.05. A type II error means claiming there is no difference between groups or no effect of a treatment, even if there is such effect. This may be avoided by doing pre study power analysis to ensure enough participants in each arm to be able to detect a possible difference between the groups. The major limitation of this study is the small number of participants. Data from BMD measurements in a comparable group of persons with MS were not available when the study was planned.

5.2.3 External validity of the results

The generalisability of the study findings depends on whether the source population is representative also of other populations. In this study all eligible persons with MS from a homogeneous and stable population in a defined geographical area where invited to participate, and the response rate was 65%. Selection bias may be a limitation in our study due to the fact that volunteers in a population based study often are more interested in health issues and may represent a healthier population than those not responding on the invitation. It may also be that the more disabled persons with MS were less likely to participate, but assuming this, the associations of physical function with BMD are rather under-than

overestimated. In addition, persons who probably represent those at highest risk of developing osteoporosis, those with medications or concomitant conditions known to be associated with secondary bone loss, were excluded from this study, thus limiting the external validity of the results. The main outcome, BMD, is a surrogate outcome for fracture risk and the trial protocol supplementing 20,000 IU vitamin D weekly which varies greatly from the routine practice in Norway, both of which reduce the validity of the results. The absence of associations with several known risk factors for low BMD [Bainbridge *et al.* 2004;Lewiecki 2005;Poole and Compston 2006;Vondracek *et al.* 2009] may be ascribed to the small number or the young age of participants in this study. Thus, negative results should not be interpreted in the sense that general risk factors are not relevant in persons with MS.

6. Implications for further research

6.1 Bone loss and vitamin D supplementation

In this phase II trial we did found that a weekly dose of 20,000 IU vitamin D₃ administrated for 96 weeks could not prevent bone loss in fully ambulatory persons with MS age 18-50 year. When the trial was planned power analysis was not performed due to lack of data on BMD change in young ambulatory persons with MS. Even though we invited all eligible persons managed by the Neurological department University Hospital in North-Norway with MS to participate in this study, we were not able to include enough participants in each treatment arm. Based on change in BMD from this study, a trial with approximately 250 persons in each treatment arm will be needed to confirm or reject the hypothesis that a weekly dose of 20,000 vitamin D for 96 weeks could prevent bone loss in persons with MS (two tailed, power 80%, risk of type I error 5% and drop out rate 10%). Adequately powered long-term studies should be performed before drawing a final conclusion.

6.2 Vitamin D levels

Optimising vitamin D status may favorably affect the disease course of MS [Correale *et al.* 2009;Smolders *et al.* 2008b;van der Mei *et al.* 2007], but required vitamin D levels are still unknown. In this study most patients obtain the recommended adequate vitamin D level (≥ 50 nmol/l) with regular sun exposure or vitamin D supplementation. The CHOLINE study (NCT01198132) and the SOLAR study (NCT01285401) and several other clinical trial with vitamin D supplementation in persons with MS are either ongoing or completed and not yet published [http://clinicaltrials.gov]. If future studies find that there is a need for higher serum

vitamin D level, we have shown that supplementation with weekly dose of 20,000 IU Vitamin D_3 brings the majority to serum vitamin D level at least ≥ 75 nmol/L. However, it is important to make regular vitamin D measurements since the individual response of vitamin D supplementation is varying.

7. Concluding remarks

In persons with MS several studies have shown that BMD decreases as disability increasingly limits physical activity. Serum levels of 25(OH)D are associated with higher BMD, and vitamin D and calcium supplementation is the basis in osteoporosis prevention and treatment. In this trial a weekly dose of 20,000 IU vitamin D₃ in 96 weeks did not affect BMD in young fully ambulatory persons with MS. However, almost 25% of the population had lower than expected BMD for sex and age, and over the study period mean BMD at the hip in the whole study population decreased significantly by 1%. This finding may indicate that MS may be a cause of secondary osteoporosis. Progressing physical handicap poses persons with MS at further risk of developing osteoporosis and at the same time increases the risk of falls and fractures. Evidence is accumulating that hypovitaminosis D may play a role in the aetiology and pathogenesis of MS, which by itself might put persons with MS at risk of low peak bone mass. Only half of the participants in this study had at study start sufficient vitamin D levels (≥ 50 nmol/L) during the winter months, but the chance of being vitamin D sufficient increased with regular sunexposure and low dose vitamin D supplementation. Supplementation with high dose vitamin D₃ weekly resulted in all participants in the treatment group being vitamin D sufficient during winter and did not cause adverse effects. Establishing decision rules for selecting individuals for BMD screening has proven difficult [Raisz 2005]. Findings in our limited sample indicate that men as well as women with MS are at increased risk of low BMD. It might be appropriate to perform BMD measurements in all patients, perhaps shortly after diagnosis, but at least when the disease affect the ambulatory function. In order to achieve adequate vitamin D levels, MS patients who have no vitamin D efficient sun exposure and low dietary vitamin D intake during the winter months should be recommended to take vitamin D supplements.

References

MSFC administration and scoring manual. MSFC administration and scoring manual. Availble at: http://www.nationalmssociety.org. Accessed 9 August 2012.

http://clinicaltrials.gov/. Assesed 15 August 2012.

Adams JS, Hewison M. Unexpected actions of vitamin D: new perspectives on the regulation of innate and adaptive immunity. Nat Clin Pract Endocrinol Metab 2008; 4: 80-90.

Adams JS, Hewison M. Update in vitamin D. J Clin Endocrinol Metab 2010; 95: 471-478.

Alonso A, Hernan MA. Temporal trends in the incidence of multiple sclerosis: a systematic review. Neurology 2008; 71: 129-135.

Altintas A, Saruhan-Direskeneli G, Benbir G, Demir M, Purisa S. The role of osteopontin: a shared pathway in the pathogenesis of multiple sclerosis and osteoporosis? J Neurol Sci 2009; 276: 41-44.

Amezcua L, Chung RH, Conti DV, Langer-Gould AM. Vitamin D levels in Hispanics with multiple sclerosis. J Neurol 2012. DOI 10.1007/s00415-012-6537-z

Ascherio A, Munger KL. Environmental risk factors for multiple sclerosis. Part II: Noninfectious factors. Ann Neurol 2007; 61: 504-513.

Ascherio A, Munger KL, Simon KC. Vitamin D and multiple sclerosis. Lancet Neurol 2010; 9: 599-612.

Aydin G, Atalar E, Keles I *et al.* Predictive value of grip strength for bone mineral density in males: site specific or systemic? Rheumatol Int 2006; 27: 125-129.

Baarnhielm M, Hedstrom AK, Kockum I *et al.* Sunlight is associated with decreased multiple sclerosis risk: no interaction with human leukocyte antigen-DRB1*15. Eur J Neurol 2012; 19: 955-962.

Bainbridge KE, Sowers M, Lin X, Harlow SD. Risk factors for low bone mineral density and the 6-year rate of bone loss among premenopausal and perimenopausal women. Osteoporos Int 2004; 15: 439-446.

Balashov KE, Olek MJ, Smith DR, Khoury SJ, Weiner HL. Seasonal variation of interferongamma production in progressive multiple sclerosis. Ann Neurol 1998; 44: 824-828.

Barnes MS, Bonham MP, Robson PJ *et al*. Assessment of 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D3 concentrations in male and female multiple sclerosis patients and control volunteers. Mult Scler 2007; 13: 670-672.

Bazelier MT, Bentzen J, Vestergaard P *et al*. The risk of fracture in incident multiple sclerosis patients: The Danish National Health Registers. Mult Scler 2012a. DOI:10.1177/1352458512442755

Bazelier MT, de VF, Bentzen J *et al.* Incidence of fractures in patients with multiple sclerosis: the Danish National Health Registers. Mult Scler 2012b; 18: 622-627.

Bazelier MT, van Staa TP, Uitdehaag BM *et al*. Risk of fractures in patients with multiple sclerosis: A population-based cohort study. Neurology 2012c; 78: 1967-1973.

Bazelier MT, van ST, Uitdehaag BM *et al*. The risk of fracture in patients with multiple sclerosis: the UK general practice research database. J Bone Miner Res 2011; 26: 2271-2279.

Ben-Nun A, Wekerle H, Cohen IR. The rapid isolation of clonable antigen-specific T lymphocyte lines capable of mediating autoimmune encephalomyelitis. Eur J Immunol 1981; 11: 195-199.

Beretich B, Beretich T. Explaining multiple sclerosis prevalence by ultraviolet exposure: a geospatial analysis. Mult Scler 2009; 15: 891-898.

Berger C, Goltzman D, Langsetmo L *et al*. Peak bone mass from longitudinal data: implications for the prevalence, pathophysiology, and diagnosis of osteoporosis. J Bone Miner Res 2010; 25: 1948-1957.

Berger C, Langsetmo L, Joseph L *et al*. Change in bone mineral density as a function of age in women and men and association with the use of antiresorptive agents. CMAJ 2008; 178: 1660-1668.

Binkley N, Krueger D, Lensmeyer G. 25-hydroxyvitamin D measurement, 2009: a review for clinicians. J Clin Densitom 2009; 12: 417-427.

Bischoff-Ferrari HA, Dietrich T, Orav EJ, Dawson-Hughes B. Positive association between 25-hydroxy vitamin D levels and bone mineral density: a population-based study of younger and older adults. Am J Med 2004; 116: 634-639.

Bischoff-Ferrari HA, Willett WC, Wong JB *et al*. Prevention of nonvertebral fractures with oral vitamin D and dose dependency: a meta-analysis of randomized controlled trials. Arch Intern Med 2009; 169: 551-561.

Bonnick S.L. Bone Densitometry in Clinical Practice 2004. Book. Humana Press Inc. Totowa, New Jersey.

Braitch M, Constantinescu CS. The role of osteopontin in experimental autoimmune encephalomyelitis (EAE) and multiple sclerosis (MS). Inflamm Allergy Drug Targets 2010; 9: 249-256.

Bruck W, Bitsch A, Kolenda H, Bruck Y, Stiefel M, Lassmann H. Inflammatory central nervous system demyelination: correlation of magnetic resonance imaging findings with lesion pathology. Ann Neurol 1997; 42: 783-793.

Brustad M, Alsaker E, Engelsen O, Aksnes L, Lund E. Vitamin D status of middle-aged women at 65-71 degrees N in relation to dietary intake and exposure to ultraviolet radiation. Public Health Nutr 2004; 7: 327-335.

Burton JM, Kimball S, Vieth R *et al*. A phase I/II dose-escalation trial of vitamin D3 and calcium in multiple sclerosis. Neurology 2010; 74: 1852-1859.

Cantorna MT, Hayes CE, DeLuca HF. 1,25-Dihydroxyvitamin D3 reversibly blocks the progression of relapsing encephalomyelitis, a model of multiple sclerosis. Proc Natl Acad Sci U S A 1996; 93: 7861-7864.

Carey JJ, Delaney MF, Love TE *et al.* Dual-energy X-ray absorptiometry diagnostic discordance between Z-scores and T-scores in young adults. J Clin Densitom 2009; 12: 11-16.

Chang IC, Chiang TI, Yeh KT, Lee H, Cheng YW. Increased serum osteopontin is a risk factor for osteoporosis in menopausal women. Osteoporos Int 2010; 21: 1401-1409.

Chapurlat RD, Garnero P, Sornay-Rendu E, Arlot ME, Claustrat B, Delmas PD. Longitudinal study of bone loss in pre- and perimenopausal women: evidence for bone loss in perimenopausal women. Osteoporos Int 2000; 11: 493-498.

Comabella M, Khoury SJ. Immunopathogenesis of multiple sclerosis. Clin Immunol 2012; 142: 2-8.

Compston A, Coles A. Multiple sclerosis. Lancet 2008; 372: 1502-1517.

Compston A, McDonald I, Noseworthy JH *et al.* McAlpine's Multiple Scleroosis. Book. 2006 4th Edition Philadelphia: Churchill Livingstone Elsevier.

Confavreux C, Hutchinson M, Hours MM, Cortinovis-Tourniaire P, Moreau T. Rate of pregnancy-related relapse in multiple sclerosis. Pregnancy in Multiple Sclerosis Group. N Engl J Med 1998; 339: 285-291.

Confavreux C, Vukusic S, Moreau T, Adeleine P. Relapses and progression of disability in multiple sclerosis. N Engl J Med 2000; 343: 1430-1438.

Correale J, Ysrraelit MC, Gaitan MI. Immunomodulatory effects of Vitamin D in multiple sclerosis. Brain 2009; 132: 1146-1160.

Correale J, Ysrraelit MC, Gaitan MI. Gender differences in 1,25 dihydroxyvitamin D3 immunomodulatory effects in multiple sclerosis patients and healthy subjects. J Immunol 2010; 185: 4948-4958.

Cosman F, Nieves J, Komar L *et al*. Fracture history and bone loss in patients with MS. Neurology 1998; 51: 1161-1165.

Cranney A, Weiler HA, O'Donnell S, Puil L. Summary of evidence-based review on vitamin D efficacy and safety in relation to bone health. Am J Clin Nutr 2008; 88: 513S-519S.

Dawson-Hughes B, Heaney RP, Holick MF, Lips P, Meunier PJ, Vieth R. Estimates of optimal vitamin D status. Osteoporos Int 2005; 16: 713-716.

de Sa JC, Airas L, Bartholome E *et al.* Symptomatic therapy in multiple sclerosis: a review for a multimodal approach in clinical practice. Ther Adv Neurol Disord 2011; 4: 139-168.

DeLuca HF. Evolution of our understanding of vitamin D. Nutr Rev 2008; 66: S73-S87.

Dhib-Jalbut S. Pathogenesis of myelin/oligodendrocyte damage in multiple sclerosis. Neurology 2007; 68: S13-S21.

Dickinson J, Perera D, van der MA *et al*. Past environmental sun exposure and risk of multiple sclerosis: a role for the Cdx-2 Vitamin D receptor variant in this interaction. Mult Scler 2009; 15: 563-570.

Diehl JW, Chiu MW. Effects of ambient sunlight and photoprotection on vitamin D status. Dermatol Ther 2010; 23: 48-60.

Donaldson LJ, Reckless IP, Scholes S, Mindell JS, Shelton NJ. The epidemiology of fractures in England. J Epidemiol Community Health 2008; 62: 174-180.

Dovio A, Perazzolo L, Osella G *et al.* Immediate fall of bone formation and transient increase of bone resorption in the course of high-dose, short-term glucocorticoid therapy in young patients with multiple sclerosis. J Clin Endocrinol Metab 2004; 89: 4923-4928.

Durazo-Arvizu RA, Dawson-Hughes B, Sempos CT *et al*. Three-phase model harmonizes estimates of the maximal suppression of parathyroid hormone by 25-hydroxyvitamin D in persons 65 years of age and older. J Nutr 2010; 140: 595-599.

Dusso AS, Brown AJ, Slatopolsky E. Vitamin D. Am J Physiol Renal Physiol 2005; 289: F8-28.

Dwyer T, van dM, I, Ponsonby AL *et al.* Melanocortin 1 receptor genotype, past environmental sun exposure, and risk of multiple sclerosis. Neurology 2008; 71: 583-589.

Dyment DA, Ebers GC, Sadovnick AD. Genetics of multiple sclerosis. Lancet Neurol 2004; 3: 104-110.

Ebers GC. Environmental factors and multiple sclerosis. Lancet Neurol 2008; 7: 268-277.

Emaus N, Berntsen GK, Joakimsen RM, Fonnebo V. Longitudinal changes in forearm bone mineral density in women and men aged 25-44 years: the Tromso study: a population-based study. Am J Epidemiol 2005; 162: 633-643.

Emaus N, Omsland TK, Ahmed LA, Grimnes G, Sneve M, Berntsen GK. Bone mineral density at the hip in Norwegian women and men--prevalence of osteoporosis depends on chosen references: the Tromso Study. Eur J Epidemiol 2009; 24: 321-328.

Embry AF, Snowdon LR, Vieth R. Vitamin D and seasonal fluctuations of gadolinium-enhancing magnetic resonance imaging lesions in multiple sclerosis. Ann Neurol 2000; 48: 271-272.

Engelsen O, Brustad M, Aksnes L, Lund E. Daily duration of vitamin D synthesis in human skin with relation to latitude, total ozone, altitude, ground cover, aerosols and cloud thickness. Photochem Photobiol 2005; 81: 1287-1290.

Eyles DW, Smith S, Kinobe R, Hewison M, McGrath JJ. Distribution of the vitamin D receptor and 1 alpha-hydroxylase in human brain. J Chem Neuroanat 2005; 29: 21-30.

Fernandes de Abreu DA, Eyles D, Feron F. Vitamin D, a neuro-immunomodulator: implications for neurodegenerative and autoimmune diseases. Psychoneuroendocrinology 2009; 34 Suppl 1: S265-S277.

Fischer JS, Rudick RA, Cutter GR, Reingold SC. The Multiple Sclerosis Functional Composite Measure (MSFC): an integrated approach to MS clinical outcome assessment. National MS Society Clinical Outcomes Assessment Task Force. Mult Scler 1999; 5: 244-250.

Formica CA, Cosman F, Nieves J, Herbert J, Lindsay R. Reduced bone mass and fat-free mass in women with multiple sclerosis: effects of ambulatory status and glucocorticoid Use. Calcif Tissue Int 1997; 61: 129-133.

Freedman DM, Dosemeci M, Alavanja MC. Mortality from multiple sclerosis and exposure to residential and occupational solar radiation: a case-control study based on death certificates. Occup Environ Med 2000; 57: 418-421.

Frohman EM, Racke MK, Raine CS. Multiple sclerosis--the plaque and its pathogenesis. N Engl J Med 2006; 354: 942-955.

Gelfand JM, Cree BA, McElroy J *et al.* Vitamin D in African Americans with multiple sclerosis. Neurology 2011; 76: 1824-1830.

Gibson JC, Summers GD. Bone health in multiple sclerosis. Osteoporos Int 2011; 22: 2935-2949.

Gillespie LD, Robertson MC, Gillespie WJ et al. Interventions for preventing falls in older people living in the community. Cochrane Database Syst Rev 2009; CD007146.

Goldacre MJ, Seagroatt V, Yeates D, ACHESON ED. Skin cancer in people with multiple sclerosis: a record linkage study. J Epidemiol Community Health 2004; 58: 142-144.

Gray O., Jolley D., Gibson K., Trojano M. Onset of relapses in multiple sclerosis: the effect of seasonal change in both the northern and southern hemisphere. Multiple Sclerosis 2009 15(suppl 2): p532-s158 (Abstract).

Grimnes G, Almaas B, Eggen AE *et al*. Effect of smoking on the serum levels of 25-hydroxyvitamin D depends on the assay employed. Eur J Endocrinol 2010; 163: 339-348.

Hagenau T, Vest R, Gissel TN *et al*. Global vitamin D levels in relation to age, gender, skin pigmentation and latitude: an ecologic meta-regression analysis. Osteoporos Int 2009; 20: 133-140.

Hammond SR, English DR, McLeod JG. The age-range of risk of developing multiple sclerosis: evidence from a migrant population in Australia. Brain 2000; 123 (Pt 5): 968-974.

Hanwell H., Vieth R, Magalhaes S., McMgowan S. Vitamin D status as a predictor of MS outcome following an initial peadiatric demyelinating event. Mult Scler 2009; 15:Suppl 2; S40 (Abstract)

Harlow SD, Crawford S, Dennerstein L, Burger HG, Mitchell ES, Sowers MF. Recommendations from a multi-study evaluation of proposed criteria for staging reproductive aging. Climacteric 2007; 10: 112-119.

Hauser SL, Weiner HL, Che M, Shapiro ME, Gilles F, Letvin NL. Prevention of experimental allergic encephalomyelitis (EAE) in the SJL/J mouse by whole body ultraviolet irradiation. J Immunol 1984; 132: 1276-1281.

Hayes CE, Donald AE. A unifying multiple sclerosis etiology linking virus infection, sunlight, and vitamin D, through viral interleukin-10. Med Hypotheses 2008; 71: 85-90.

Heaney RP, Abrams S, Dawson-Hughes B *et al.* Peak bone mass. Osteoporos Int 2000; 11: 985-1009.

Heaney RP, Dowell MS, Hale CA, Bendich A. Calcium absorption varies within the reference range for serum 25-hydroxyvitamin D. J Am Coll Nutr 2003; 22: 142-146.

Heaney RP, Recker RR, Grote J, Horst RL, Armas LA. Vitamin D(3) is more potent than vitamin D(2) in humans. J Clin Endocrinol Metab 2011; 96: E447-E452.

Hearn AP, Silber E. Osteoporosis in multiple sclerosis. Mult Scler 2010; 16: 1031-1043.

Henry MJ, Pasco JA, Korn S, Gibson JE, Kotowicz MA, Nicholson GC. Bone mineral density reference ranges for Australian men: Geelong Osteoporosis Study. Osteoporos Int 2010; 21: 909-917.

Herbert J J-SKRHKLLNGAeal. The timed tandem walk is a more informative measure of mild impairment in patients with multiple sclerosis than the 25-foot walk. Mult Scler 2006; 12:S87 (Abstract)

Hiremath GS, Cettomai D, Baynes M *et al*. Vitamin D status and effect of low-dose cholecalciferol and high-dose ergocalciferol supplementation in multiple sclerosis. Mult Scler 2009; 15: 735-740.

Hjartaker A, Andersen LF, Lund E. Comparison of diet measures from a food-frequency questionnaire with measures from repeated 24-hour dietary recalls. The Norwegian Women and Cancer Study. Public Health Nutr 2007; 10: 1094-1103.

Hofbauer LC, Hamann C, Ebeling PR. Approach to the patient with secondary osteoporosis. Eur J Endocrinol 2010; 162: 1009-1020.

Holick MF. Vitamin D deficiency. N Engl J Med 2007; 357: 266-281.

Holick MF, Binkley NC, Bischoff-Ferrari HA *et al.* Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab 2011; 96: 1911-1930.

Hollis BW. Circulating 25-hydroxyvitamin D levels indicative of vitamin D sufficiency: implications for establishing a new effective dietary intake recommendation for vitamin D. J Nutr 2005; 135: 317-322.

Hollis BW. Assessment of vitamin D status and definition of a normal circulating range of 25-hydroxyvitamin D. Curr Opin Endocrinol Diabetes Obes 2008; 15: 489-494.

Holmoy T. Immunopathogenesis of multiple sclerosis: concepts and controversies. Acta Neurol Scand Suppl 2007; 187: 39-45.

Holmoy T. Vitamin D status modulates the immune response to Epstein Barr virus: Synergistic effect of risk factors in multiple sclerosis. Med Hypotheses 2008; 70: 66-69.

Holmoy T, Moen SM, Gundersen TA *et al.* 25-hydroxyvitamin D in cerebrospinal fluid during relapse and remission of multiple sclerosis. Mult Scler 2009; 15: 1280-1285.

Islam T, Gauderman WJ, Cozen W, Hamilton AS, Burnett ME, Mack TM. Differential twin concordance for multiple sclerosis by latitude of birthplace. Ann Neurol 2006; 60: 56-64.

Islam T, Gauderman WJ, Cozen W, Mack TM. Childhood sun exposure influences risk of multiple sclerosis in monozygotic twins. Neurology 2007; 69: 381-388.

Jones G. Pharmacokinetics of vitamin D toxicity. Am J Clin Nutr 2008; 88: 582S-586S.

Kampman MT, Brustad M. Vitamin D: a candidate for the environmental effect in multiple sclerosis - observations from Norway. Neuroepidemiology 2008; 30: 140-146.

Kampman MT, Eriksen EF, Holmoy T. Multiple sclerosis, a cause of secondary osteoporosis? What is the evidence and what are the clinical implications? Acta Neurol Scand Suppl 2011; 44-49.

Kampman MT, Steffensen LH, Mellgren SI, Jorgensen L. Effect of vitamin D3 supplementation on relapses, disease progression, and measures of function in persons with multiple sclerosis: exploratory outcomes from a double-blind randomised controlled trial. Mult Scler 2012; 18: 1144-1151.

Kampman MT, Wilsgaard T, Mellgren SI. Outdoor activities and diet in childhood and adolescence relate to MS risk above the Arctic Circle. J Neurol 2007; 254: 471-477.

Kanis JA. Diagnosis of osteoporosis and assessment of fracture risk. Lancet 2002; 359: 1929-1936.

Kappos L, Radue EW, O'Connor P *et al*. A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis. N Engl J Med 2010; 362: 387-401.

Killestein J, Rep MH, Meilof JF *et al.* Seasonal variation in immune measurements and MRI markers of disease activity in MS. Neurology 2002; 58: 1077-1080.

Kimball SM, Ursell MR, O'Connor P, Vieth R. Safety of vitamin D3 in adults with multiple sclerosis. Am J Clin Nutr 2007; 86: 645-651.

Knippenberg S, Bol Y, Damoiseaux J, Hupperts R, Smolders J. Vitamin D status in patients with MS is negatively correlated with depression, but not with fatigue. Acta Neurol Scand 2011; 124: 171-175.

Kong YY, Boyle WJ, Penninger JM. Osteoprotegerin ligand: a regulator of immune responses and bone physiology. Immunol Today 2000; 21: 495-502.

Kurban S, Akpinar Z, Mehmetoglu I. Receptor activator of nuclear factor kappaB ligand (RANKL) and osteoprotegerin levels in multiple sclerosis. Mult Scler 2008; 14: 431-432.

Kurtzke JF. Rating neurologic impairment in multiple sclerosis: an expanded disability status scale (EDSS). Neurology 1983; 33: 1444-1452.

Lange NE, Litonjua A, Hawrylowicz CM, Weiss S. Vitamin D, the immune system and asthma. Expert Rev Clin Immunol 2009; 5: 693-702.

Lemire JM, Archer DC. 1,25-dihydroxyvitamin D3 prevents the in vivo induction of murine experimental autoimmune encephalomyelitis. J Clin Invest 1991; 87: 1103-1107.

Leslie WD, Adler RA, El-Hajj FG *et al.* Application of the 1994 WHO classification to populations other than postmenopausal Caucasian women: the 2005 ISCD Official Positions. J Clin Densitom 2006; 9: 22-30.

Lewiecki EM. Premenopausal bone health assessment. Curr Rheumatol Rep 2005; 7: 46-52.

Logan WC, Jr., Sloane R, Lyles KW, Goldstein B, Hoenig HM. Incidence of fractures in a cohort of veterans with chronic multiple sclerosis or traumatic spinal cord injury. Arch Phys Med Rehabil 2008; 89: 237-243.

Loken-Amsrud KI, Holmoy T, Bakke SJ *et al.* Vitamin D and disease activity in multiple sclerosis before and during interferon-beta treatment. Neurology 2012; 79: 267-273.

Lonergan R, Kinsella K, Fitzpatrick P *et al.* Multiple sclerosis prevalence in Ireland: relationship to vitamin D status and HLA genotype. J Neurol Neurosurg Psychiatry 2011; 82: 317-322.

Lucas RM, Ponsonby AL. Considering the potential benefits as well as adverse effects of sun exposure: can all the potential benefits be provided by oral vitamin D supplementation? Prog Biophys Mol Biol 2006; 92: 140-149.

Lucas RM, Ponsonby AL, Dear K *et al.* Sun exposure and vitamin D are independent risk factors for CNS demyelination. Neurology 2011; 76: 540-548.

Macdonald HM, Mavroeidi A, Barr RJ, Black AJ, Fraser WD, Reid DM. Vitamin D status in postmenopausal women living at higher latitudes in the UK in relation to bone health, overweight, sunlight exposure and dietary vitamin D. Bone 2008; 42: 996-1003.

Mahon BD, Gordon SA, Cruz J, Cosman F, Cantorna MT. Cytokine profile in patients with multiple sclerosis following vitamin D supplementation. J Neuroimmunol 2003; 134: 128-132.

Marrie RA. Environmental risk factors in multiple sclerosis aetiology. Lancet Neurol 2004; 3: 709-718.

McDonald WI, Compston A, Edan G *et al*. Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the diagnosis of multiple sclerosis. Ann Neurol 2001; 50: 121-127.

McDowell TY, Amr S, Culpepper WJ *et al*. Sun exposure, vitamin D intake and progression to disability among veterans with progressive multiple sclerosis. Neuroepidemiology 2011; 37: 52-57.

Meyer HE, Brunvand L, Brustad M, Holvik K, Johansson L, Paulsen JE. [Report IS-1408: Measures to ensure a good vitamin D status in the population]. Oslo: Norwegian Directorate of Health 2006.

Michels KB, Willett WC, Graubard BI *et al.* A longitudinal study of infant feeding and obesity throughout life course. Int J Obes (Lond) 2007; 31: 1078-1085.

Miller DH, Leary SM. Primary-progressive multiple sclerosis. Lancet Neurol 2007; 6: 903-912.

Miller E. Multiple sclerosis. Adv Exp Med Biol 2012; 724: 222-238.

Mithal A, Wahl DA, Bonjour JP *et al*. Global vitamin D status and determinants of hypovitaminosis D. Osteoporos Int 2009; 20: 1807-1820.

Moen SM, Celius EG, Nordsletten L, Holmoy T. Fractures and falls in patients with newly diagnosed clinically isolated syndrome and multiple sclerosis. Acta Neurol Scand Suppl 2011a; 79-82.

Moen SM, Celius EG, Sandvik L, Nordsletten L, Eriksen EF, Holmoy T. Low bone mass in newly diagnosed multiple sclerosis and clinically isolated syndrome. Neurology 2011b; 77: 151-157.

Mohr DC, Hart SL, Julian L, Cox D, Pelletier D. Association between stressful life events and exacerbation in multiple sclerosis: a meta-analysis. BMJ 2004; 328: 731.

Munger KL, Levin LI, Hollis BW, Howard NS, Ascherio A. Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. JAMA 2006; 296: 2832-2838.

Munger KL, Zhang SM, O'Reilly E *et al*. Vitamin D intake and incidence of multiple sclerosis. Neurology 2004; 62: 60-65.

Myhr KM, Mellgren SI. Corticosteroids in the treatment of multiple sclerosis. Acta Neurol Scand Suppl 2009; 73-80.

Nashold FE, Spach KM, Spanier JA, Hayes CE. Estrogen controls vitamin D3-mediated resistance to experimental autoimmune encephalomyelitis by controlling vitamin D3 metabolism and receptor expression. J Immunol 2009; 183: 3672-3681.

Neer RM. Bone loss across the menopausal transition. Ann N Y Acad Sci 2010; 1192: 66-71.

Nieves J, Cosman F, Herbert J, Shen V, Lindsay R. High prevalence of vitamin D deficiency and reduced bone mass in multiple sclerosis. Neurology 1994; 44: 1687-1692.

Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. N Engl J Med 2000; 343: 938-952.

O'Mahony L, Stepien M, Gibney MJ, Nugent AP, Brennan L. The potential role of vitamin D enhanced foods in improving vitamin D status. Nutrients 2011; 3: 1023-1041.

Orton SM, Herrera BM, Yee IM *et al.* Sex ratio of multiple sclerosis in Canada: a longitudinal study. Lancet Neurol 2006; 5: 932-936.

Orton SM, Morris AP, Herrera BM *et al*. Evidence for genetic regulation of vitamin D status in twins with multiple sclerosis. Am J Clin Nutr 2008; 88: 441-447.

Ozgocmen S, Bulut S, Ilhan N, Gulkesen A, Ardicoglu O, Ozkan Y. Vitamin D deficiency and reduced bone mineral density in multiple sclerosis: effect of ambulatory status and functional capacity. J Bone Miner Metab 2005; 23: 309-313.

Pearce SH, Cheetham TD. Diagnosis and management of vitamin D deficiency. BMJ 2010; 340: b5664.

Pelletier D, Hafler DA. Fingolimod for multiple sclerosis. N Engl J Med 2012; 366: 339-347.

Peterlik M, Cross HS. Vitamin D and calcium deficits predispose for multiple chronic diseases. Eur J Clin Invest 2005; 35: 290-304.

Pierrot-Deseilligny C. Clinical implications of a possible role of vitamin D in multiple sclerosis. J Neurol 2009; 256: 1468-1479.

Polman CH, O'Connor PW, Havrdova E *et al*. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. N Engl J Med 2006; 354: 899-910.

Polman CH, Reingold SC, Banwell B *et al.* Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. Ann Neurol 2011; 69: 292-302.

Poole KE, Compston JE. Osteoporosis and its management. BMJ 2006; 333: 1251-1256.

Pugliatti M, Harbo HF, Holmoy T *et al*. Environmental risk factors in multiple sclerosis. Acta Neurol Scand Suppl 2008a; 188: 34-40.

Pugliatti M, Riise T, Nortvedt MW *et al.* Self-perceived physical functioning and health status among fully ambulatory multiple sclerosis patients. J Neurol 2008b; 255: 157-162.

Pugliatti M, Rosati G, Carton H *et al*. The epidemiology of multiple sclerosis in Europe. Eur J Neurol 2006; 13: 700-722.

Pugliatti M, Sotgiu S, Rosati G. The worldwide prevalence of multiple sclerosis. Clin Neurol Neurosurg 2002; 104: 182-191.

Raisz LG. Pathogenesis of osteoporosis: concepts, conflicts, and prospects. J Clin Invest 2005; 115: 3318-3325.

Ramagopalan SV, Knight JC, Ebers GC. Multiple sclerosis and the major histocompatibility complex. Curr Opin Neurol 2009; 22: 219-225.

Ravnborg M, Sorensen PS, Andersson M *et al.* Methylprednisolone in combination with interferon beta-1a for relapsing-remitting multiple sclerosis (MECOMBIN study): a multicentre, double-blind, randomised, placebo-controlled, parallel-group trial. Lancet Neurol 2010; 9: 672-680.

Recker R, Lappe J, Davies K, Heaney R. Characterization of perimenopausal bone loss: a prospective study. J Bone Miner Res 2000; 15: 1965-1973.

Rosen CJ. Clinical practice. Vitamin D insufficiency. N Engl J Med 2011; 364: 248-254.

Ross AC, Manson JE, Abrams SA *et al*. The 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of Medicine: what clinicians need to know. J Clin Endocrinol Metab 2011; 96: 53-58.

Runia TF, Hop WC, de Rijke YB, Buljevac D, Hintzen RQ. Lower serum vitamin D levels are associated with a higher relapse risk in multiple sclerosis. Neurology 2012; 79: 261-266.

Sambrook P, Cooper C. Osteoporosis. Lancet 2006; 367: 2010-2018.

Sandhu SK, Hampson G. The pathogenesis, diagnosis, investigation and management of osteoporosis. J Clin Pathol 2011; 64: 1042-1050.

Schwid SR, Goodman AD, Puzas JE, McDermott MP, Mattson DH. Sporadic corticosteroid pulses and osteoporosis in multiple sclerosis. Arch Neurol 1996; 53: 753-757.

Seeman E, Delmas PD. Bone quality--the material and structural basis of bone strength and fragility. N Engl J Med 2006; 354: 2250-2261.

Sibley WA, Bamford CR, Clark K, Smith MS, Laguna JF. A prospective study of physical trauma and multiple sclerosis. J Neurol Neurosurg Psychiatry 1991; 54: 584-589.

Simpson S Jr, Taylor B, Blizzard L *et al*. Higher 25-hydroxyvitamin D is associated with lower relapse risk in multiple sclerosis. Ann Neurol 2010; 68: 193-203.

Sioka C, Papakonstantinou S, Fotopoulos A *et al*. Bone mineral density in ambulatory patients with multiple sclerosis. Neurol Sci 2011; 32: 819-824.

Siris ES, Chen YT, Abbott TA *et al.* Bone mineral density thresholds for pharmacological intervention to prevent fractures. Arch Intern Med 2004; 164: 1108-1112.

Sloka S, Silva C, Pryse-Phillips W, Patten S, Metz L, Yong VW. A quantitative analysis of suspected environmental causes of MS. Can J Neurol Sci 2011; 38: 98-105.

Smolders J, Damoiseaux J, Menheere P, Hupperts R. Vitamin D as an immune modulator in multiple sclerosis, a review. J Neuroimmunol 2008a; 194: 7-17.

Smolders J, Menheere P, Kessels A, Damoiseaux J, Hupperts R. Association of vitamin D metabolite levels with relapse rate and disability in multiple sclerosis. Mult Scler 2008b; 14: 1220-1224.

Smolders J, Peelen E, Thewissen M *et al*. The relevance of vitamin D receptor gene polymorphisms for vitamin D research in multiple sclerosis. Autoimmun Rev 2009a; 8: 621-626.

Smolders J, Thewissen M, Peelen E *et al*. Vitamin D status is positively correlated with regulatory T cell function in patients with multiple sclerosis. PLoS One 2009b; 4: e6635.

Soilu-Hanninen M, Airas L, Mononen I, Heikkila A, Viljanen M, Hanninen A. 25-Hydroxyvitamin D levels in serum at the onset of multiple sclerosis. Mult Scler 2005; 11: 266-271.

Soilu-Hanninen M, Aivo J, Lindstrom BM *et al*. A randomised, double blind, placebo controlled trial with vitamin D3 as an add on treatment to interferon beta-1b in patients with multiple sclerosis. J Neurol Neurosurg Psychiatry 2012; 83: 565-571.

Soilu-Hanninen M, Laaksonen M, Laitinen I, Eralinna JP, Lilius EM, Mononen I. A longitudinal study of serum 25-hydroxyvitamin D and intact parathyroid hormone levels indicate the importance of vitamin D and calcium homeostasis regulation in multiple sclerosis. J Neurol Neurosurg Psychiatry 2008; 79: 152-157.

Sorensen PS, Mellgren SI, Svenningsson A *et al.* NORdic trial of oral Methylprednisolone as add-on therapy to Interferon beta-1a for treatment of relapsing-remitting Multiple Sclerosis (NORMIMS study): a randomised, placebo-controlled trial. Lancet Neurol 2009; 8: 519-529.

Spach KM, Hayes CE. Vitamin D3 confers protection from autoimmune encephalomyelitis only in female mice. J Immunol 2005; 175: 4119-4126.

Stein MS, Liu Y, Gray OM *et al*. A randomized trial of high-dose vitamin D2 in relapsing-remitting multiple sclerosis. Neurology 2011; 77: 1611-1618.

Stepan JJ, Havrdova E, Tyblova M *et al.* Markers of bone remodeling predict rate of bone loss in multiple sclerosis patients treated with low dose glucocorticoids. Clin Chim Acta 2004; 348: 147-154.

Styrkarsdottir U, Halldorsson BV, Gretarsdottir S *et al*. New sequence variants associated with bone mineral density. Nat Genet 2009; 41: 15-17.

Tang BM, Eslick GD, Nowson C, Smith C, Bensoussan A. Use of calcium or calcium in combination with vitamin D supplementation to prevent fractures and bone loss in people aged 50 years and older: a meta-analysis. Lancet 2007; 370: 657-666.

Terzi T, Terzi M, Tander B, Canturk F, Onar M. Changes in bone mineral density and bone metabolism markers in premenopausal women with multiple sclerosis and the relationship to clinical variables. J Clin Neurosci 2010; 17: 1260-1264.

Thacker EL, Mirzaei F, Ascherio A. Infectious mononucleosis and risk for multiple sclerosis: a meta-analysis. Ann Neurol 2006; 59: 499-503.

Tremlett H, Paty D, Devonshire V. Disability progression in multiple sclerosis is slower than previously reported. Neurology 2006; 66: 172-177.

Tremlett H, van dM, I, Pittas F *et al.* Monthly ambient sunlight, infections and relapse rates in multiple sclerosis. Neuroepidemiology 2008; 31: 271-279.

Tremlett H, Zhao Y, Rieckmann P, Hutchinson M. New perspectives in the natural history of multiple sclerosis. Neurology 2010; 74: 2004-2015.

Triantafyllou N, Lambrinoudaki I, Thoda P *et al.* Lack of association between vitamin D levels and bone mineral density in patients with multiple sclerosis. J Neurol Sci 2012; 313: 137-141.

Troiano RA, Jotkowitz A, Cook SD, Bansil S, Zito G. Rate and types of fractures in corticosteroid-treated multiple sclerosis patients. Neurology 1992; 42: 1389-1391.

Tuzun S, Altintas A, Karacan I, Tangurek S, Saip S, Siva A. Bone status in multiple sclerosis: beyond corticosteroids. Mult Scler 2003; 9: 600-604.

Vainionpaa A, Korpelainen R, Leppaluoto J, Jamsa T. Effects of high-impact exercise on bone mineral density: a randomized controlled trial in premenopausal women. Osteoporos Int 2005; 16: 191-197.

van dM, I, Blizzard L, Ponsonby AL, Dwyer T. Validity and reliability of adult recall of past sun exposure in a case-control study of multiple sclerosis. Cancer Epidemiol Biomarkers Prev 2006; 15: 1538-1544.

van dM, I, Ponsonby AL, Blizzard L, Dwyer T. Regional variation in multiple sclerosis prevalence in Australia and its association with ambient ultraviolet radiation. Neuroepidemiology 2001; 20: 168-174.

van dM, I, Ponsonby AL, Dwyer T *et al*. Past exposure to sun, skin phenotype, and risk of multiple sclerosis: case-control study. BMJ 2003; 327: 316.

van dM, I, Ponsonby AL, Dwyer T *et al.* Vitamin D levels in people with multiple sclerosis and community controls in Tasmania, Australia. J Neurol 2007; 254: 581-590.

Van Amerongen BM, Dijkstra CD, Lips P, Polman CH. Multiple sclerosis and vitamin D: an update. Eur J Clin Nutr 2004; 58: 1095-1109.

Vatne A, Mygland A, Ljostad U. Multiple sclerosis in Vest-Agder County, Norway. Acta Neurol Scand 2011; 123: 396-399.

Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4+CD25+ regulatory T cells in patients with multiple sclerosis. J Exp Med 2004; 199: 971-979.

Vogt MH, Lopatinskaya L, Smits M, Polman CH, Nagelkerken L. Elevated osteopontin levels in active relapsing-remitting multiple sclerosis. Ann Neurol 2003; 53: 819-822.

Vondracek SF, Hansen LB, McDermott MT. Osteoporosis risk in premenopausal women. Pharmacotherapy 2009; 29: 305-317.

Vukusic S, Van B, V, Gosselin S, Confavreux C. Regional variations in the prevalence of multiple sclerosis in French farmers. J Neurol Neurosurg Psychiatry 2007; 78: 707-709.

Weinstock-Guttman B, Gallagher E, Baier M *et al*. Risk of bone loss in men with multiple sclerosis. Mult Scler 2004; 10: 170-175.

Weinstock-Guttman B, Hong J, Santos R *et al.* Interferon-beta modulates bone-associated cytokines and osteoclast precursor activity in multiple sclerosis patients. Mult Scler 2006; 12: 541-550.

Weinstock-Guttman B, Zivadinov R, Ramanathan M. Inter-dependence of vitamin D levels with serum lipid profiles in multiple sclerosis. J Neurol Sci 2011; 311: 86-91.

Woolmore JA, Stone M, Pye EM *et al.* Studies of associations between disability in multiple sclerosis, skin type, gender and ultraviolet radiation. Mult Scler 2007; 13: 369-375.

Writing group for the ISCD position development conference. Diagnosis of osteoporosis in men, premenopausal women, and children. J Clin Densitom 2004; 7: 17-26.

Yildiz M, Tettenborn B, Putzki N. Vitamin D levels in Swiss multiple sclerosis patients. Swiss Med Wkly 2011; 141: w13192.

Zorzon M, Zivadinov R, Locatelli L *et al*. Long-term effects of intravenous high dose methylprednisolone pulses on bone mineral density in patients with multiple sclerosis. Eur J Neurol 2005; 12: 550-556.



Paper II

Paper III

Appendix A

"Hva spiser du?" Norwegian version

ID			

HVA SPISER DU?

Dette spørreskjemaet inneholder nokså detaljerte spørsmål om ditt kosthold. Vi er klar over at kostholdet varierer fra dag til dag og i løpet av året. Prøv derfor å tenke deg et "gjennomsnitt" av dine spisevaner. Ha det siste året i tankene når du fyller ut. Er du usikker, ber vi deg om å krysse av for det alternativet som passer best, selv om det kanskje ikke passer helt.

Det er viktig at du plasserer kryssene tydelig i boksene. Du må gjerne bruke blyant og viskelær. Bruker du kulepenn og krysser feil, fargelegg hele boksen som er feil og sett kryss i den rette boksen.

Alle opplysningene vil bli behandlet fortrolig.

Kosthold	Drikke	
Påvirker noen av følgende forhold kostholdet ditt? (sett gjerne flere kryss)	Hvor mange glass melk type? (Sett ett kryss pr. linje)	drikker du vanligvis av hve
☐ Er vegetarianer/veganer ☐ Har bulimi ☐ Spiser ikke norsk kost til daglig ☐ Prøver å gå ☐ Har allergi/intoleranse ☐ ned i vekt ☐ Lav glykemisk ☐ Har anoreksi ☐ mat Vi er interessert i å få kjennskap til hvordan kostholdet ditt er vanligvis. Kryss av for hvert spørsmål om hvor ofte du i gjennomsnitt siste året har brukt den aktuelle matvaren, og hvor mye du pleier å spise/drikke hver gang.	aldri/ sjelden Helmelk (søt, sur) Lettmelk (søt, sur) Ekstra lettmelk Skummet (søt, sur)	1-4 pr. 5-6 pr. 1 pr. 2-3 pr. 4+ uke uke dag dag pr.dag

Hvor mange kopper kaffe/te drikker du vanligvis av hver sort? (Sett ett kryss for hver linje)	På hvor mange brødskiver bruker du? (Sett ett kryss pr. linje)
aldri/ 1-6 pr. 1 pr. 2-3 pr. 4-5 pr. 6-7 pr. 8+ sjelden uke dag dag dag dag pr.dag	Aldri/ 1-3 pr. 4-6 pr. 1 pr. 2-3 pr. 4+ sjelden uke uke dag dag pr.dag
Kokekaffe	Syltetoy
Traktekaffe	Brunost, helfet
Pulverkaffe	Brunost,
Svart te	halvfet/mager
Grønn te	Hvitost, helfet
Bruker du følgende i kaffe eller te: Kaffe Te	Hvitost, halvfet/mager
Sukker (ikke kunstig søtstoff)	Kjøttpålegg,
Melk eller flote ☐Ja ☐ Nei ☐ Ja ☐ Nei	Leverpostei
Hvor mange glass vann drikker du vanligvis? (Sett ett kryss for hver linje)	italiensk o.l.
aldri/ 1-6 pr. 1 pr. 2-3 pr. 4-5 pr. 6-7 pr. 8+ sjelden uke dag dag dag dag pr.dag	På hvor mange brødskiver <u>pr. uke</u> har du i gjennomsnitt siste året spist? (Sett ett kryss pr. linje)
Springvann/	Aldri/ 1 2-3 4-6 7-9 10+
Hvor mange glass appelsinjuice, saft og brus drikker	sjelden pr.uke pr.uke pr.uke pr.uke pr.uke Makrell i tomat,
du vanligvis? (Sett ett kryss for hver linje)	røkt makrell Kaviar
aldri/ 1-3 pr. 4-6 pr. 1 pr. 2-3 pr. 4+ pr. sjelden uke uke dag dag dag	Sild/Ansjos
Appelsinjuice	Laks (gravet/rokt)
Annen juice	Annet fiskepålegg
Saft/brus med sukker	Hva slags fett bruker du vanligvis <u>på brødet?</u> (Sett gjerne flere kryss)
Saft/brus sukkerfri	☐ Bruker ikke fett på brødet
Yoghurt/kornblanding	☐ Smør
Hvor ofte spiser du yoghurt (1 beger)? (Sett ett kryss)	☐ Hard margarin (f. eks. Per, Melange) ☐ Myk margarin (f. eks. Soft, Vita, Solsikke)
☐ Aldri/sjelden ☐ 1 pr. uke	☐ Smørblandet margarin (f.eks. Bremyk)
2-3 pr. uke 4+ pr. uke	Brelett
Hvor ofte spiser du kornblanding, havregryn eller	☐ Lettmargarin (f. eks. Soft light, Letta, Vita Lett) ☐ Middels lett margarin (f. eks. Olivero, Omega)
müsli? (Sett ett kryss) ☐ Aldri/sjelden ☐ 1-3 pr. uke	
☐ Aldri/sjelden ☐ 1-3 pr. uke ☐ 4-6 pr. uke ☐ 1 pr. dag	Dersom du bruker fett på brødet, hvor tykt lag pleier
Brødmat	du å smøre på? (En kuvertpakke med margarin veier 12 gram). (Sett ett kryss)
Hvor mange skiver brød/rundstykker og knekkebrød/	Skrapet (3 g) Tynt lag (5 g)
skonrokker spiser du vanligvis? (1/2 rundstykke = 1 brødskive) (Sett ett kryss for hver linje)	☐ Godt dekket (8 g) ☐ Tykt lag (12 g)
aldri/ 1-4 pr. 5-7 pr. 2-3 pr. 4-5 pr. 6+ sjelden uke uke dag dag pr.	Frukt og grønnsaker
Grovt brød	Hvor ofte spiser du frukt? (Sett ett kryss pr. linje)
Kneipp/halvfint	aldri/ 1-3 1 2-4 5-6 1 2+
Fint brød/baguett	sjelden pr.mnd. pr.uke pr.uke pr.dag pr. dag
Knekkebrød o.l.	Epler/pærer
I neste spalte er det spørsmål om bruk av ulike	Appelsiner o.l.
påleggstyper. Vi spør om hvor mange brødskiver med det aktuelle pålegget du pleier å spise. Dersom du	Bananer
også bruker matvarene i andre sammenhenger enn til brød (f. eks. til vafler, frokostblandinger, grøt), ber vi	Annen frukt
om at du tar med dette når du besvarer spørsmålene.	

Hvor ofte spiser du ulike typer grønnsaker? (Sett ett kryss pr. linje)				
aldri/ 1-3 1 2 3 4-5 6-7 sjelden pr.mnd. pr.uke pr.uke pr.uke pr.uke pr.uke pr.uke pr.u	Vi vil gjerne vite hvor ofte du pleier å spise fisk, og ber deg fylle ut spørsmålene om fiskeforbruk så godt du kan. Tilgangen på fisk kan variere gjennom året. Vær vennlig å markere i hvilke årstider du spiser de ulike fiskeslagene.			
Kålrot	aldri/ like mye vinter vår sommer host sjelden hele året			
Brokkoli/blomkål	Torsk, sei, hyse, lyr			
Løk	Sild			
Andre grønn- saker □ □ □ □ □ □	Annen fisk			
For de grønnsakene du spiser, kryss av for hvor mye du spiser hver gang. (Sett ett kryss for hver sort)	Med tanke på de periodene av året der du spiser fi <u>s</u> k, hvor ofte pleier du å spise følgende til <u>middag?</u> (Sett ett kryss pr. linje)			
	sjelden pr. mnd. pr. uke pr. uke			
Kål	Kokt torsk,			
Kålrot _ 1/2 dl _ 1 dl _ 1 1/2 dl _ 2+ dl Brokkoli/	Stekt torsk,			
blomkål 1-2 buketter 3-4 buketter 5+ buketter Blandet salat 1 dl 2 dl 3 dl 4+ dl	Steinbit,			
Tomat 1/4 stk 1/2 stk 1stk 2+ stk	Laks, ørret			
Grønnsak-	Makrell			
blanding 1/2 dl 1 dl 2 dl 3+ dl	Sild			
Hvor mange poteter spiser du vanligvis (kokte, stekte, mos)? (Sett ett kryss)	Annen fisk			
Spiser ikke/spiser sjelden poteter 1-4 pr. uke 5-6 pr. uke 1 pr. dag 2 pr. dag	Dersom du spiser fisk, hvor mye spiser du vanligvis pr. gang? (1 skive/stykke = 150 gram)			
3 pr. dag 4+ pr. dag	Kokt fisk (skive)			
Ris, spaghetti, grøt, suppe Hvor ofte bruker du ris og spaghetti/makaroni?	Stekt fisk (stykke)			
(Sett ett kryss pr. linje) aldri/ 1-3 pr. 1 pr. 2 pr. 3+ sjelden mnd. uke uke pr.uke	Hvor mange ganger pr. år spiser du fiskeinnmat? (Sett ett kryss pr. linje)			
Ris	0 1-3 4-6 7-9 10+			
Hvor ofte spiser du grøt ? (Sett ett kryss pr. linje)	Rogn			
aldri/ 1 pr. 2-3 pr. 1 pr. 2-6 1+ sjelden mnd. mnd. uke pr. uke pr. dag	Dersom du spiser fiskelever, hvor mange spise- skjeer pleier du å spise hver gang? (Sett ett kryss)			
Risengrynsgrøt	☐ 1 ☐ 2 ☐ 3-4 ☐ 5-6 ☐ 7+			
Hvor ofte spiser du suppe?	Hvor ofte bruker du følgende typer fiskemat? (Sett ett kryss pr. linje)			
(Sett ett kryss pr. linje) aldri/ 1-3 pr. 1 pr. 2 pr. 3+ sjelden mnd. uke uke pr.uke	aldri/ 1 pr. 2-3 pr. 1 pr. 2+ sjelden mnd. mnd. uke pr. uke			
Som hovedrett	Fiskekaker/pudding/boller			

Hvor stor mengde pleier du vanligvis å spise av de ulike rettene? (Sett ett kryss for hver linje)	Dersom du spiser følgende retter, oppgi mengden du vanligvis spiser: (Sett ett kryss for hver linje)
Fiskekaker/pudding/boller (stk.)	Steik (skiver) 1 2 3 4 5+ Koteletter(stk.) 1/2 1 1,5 2+
Plukkfisk, fiskegrateng (dl)	Kjøttkaker,
Frityrfisk, fiskepinner (stk.) 1-2 3-4 5-6 7+	karbonader (stk.) 1 2 3 4+
	Pølser (stk. à 150g) 1/2 1 1,5 2+
l tillegg til informasjon om fiskeforbruk er det viktig å få kartlagt hvilket tilbehør som blir servert <u>til fisk.</u> Hvor ofte bruker du følgende til fisk?	Gryterett, lapskaus (dl)
aldri/ 1 pr. 2-3 pr. 1 pr. 2+ sjelden mnd. mnd. uke pr. uke	(stykke à 100 g) 1 2 3 4+
Smeltet/fast smør	Hvilke sauser bruker du til kjøttretter og pastaretter?
Smeltet/fast margarin/fett	(Sett ett kryss pr. linje) aldri/ 1 pr. 2-3 pr. 1 pr. 2+
Seterrømme (35%)	sjelden mnd. mnd. uke pr. uke
Lettrømme (20%)	Brun saus
Saus med fett (hvit/brun)	Sjysaus
Saus uten fett (hvit/brun)	Tomatsaus
	Saus med fløte/rømme
For de ulike typene tilbehør du bruker <u>til fisk,</u> vær vennlig å kryss av for hvor mye du vanligvis pleier å spise.	Hvor mye bruker du vanligvis av disse sausene?
	Brun saus (dl)
Smeltet/	Sjysaus (dl)
fast smør (ss)	Tomatsaus (dl) 1/4 1/2 3/4 1 2+
Smeltet/	Saus med fløte/
Seterramme (cc)	
Seterrømme (ss) 📗 1/2 📗 1 📗 2 📗 3 📗 4+	Andre matvarer
Lettrømme (ss)	
	Andre matvarer Hvor mange egg spiser du vanligvis i løpet <u>av en</u> <u>uke</u> ?(stekte, kokte, eggerøre, omelett) (Sett ett kryss)
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet <u>av en</u>
Lettromme (ss)	Hvor mange egg spiser du vanligvis i løpet <u>av en uke</u> ?(stekte, kokte, eggerøre, omelett) (Sett ett kryss)
Lettromme (ss)	Hvor mange egg spiser du vanligvis i løpet <u>av en uke</u> ?(stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0
Lettromme (ss)	Hvor mange egg spiser du vanligvis i løpet <u>av en uke</u> ?(stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0
Lettromme (ss)	Hvor mange egg spiser du vanligvis i løpet <u>av en uke</u> ?(stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0
Lettromme (ss)	Hvor mange egg spiser du vanligvis i løpet <u>av en uke</u> ?(stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0 1 2 3-4 5-6 7+ Hvor ofte spiser du iskrem? (til dessert, Krone-is osv.) Sett ett kryss for hvor ofte du spiser iskrem om sommeren, og ett kryss for resten av året
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (sett ett kryss) 0
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke?(stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0 1 2 3-4 5-6 7+ Hvor ofte spiser du iskrem? (till dessert, Krone-is osv.) Sett ett kryss for hvor ofte du spiser iskrem om sommeren, og ett kryss for resten av året aldri/ 1 pr. 2-3 pr. 1 pr. 2+ sjelden mnd. mnd. uke pr.uke Om sommeren
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0 1 2 3-4 5-6 7+ Hvor ofte spiser du iskrem? (till dessert, Krone-is osv.) Sett ett kryss for hvor ofte du spiser iskrem om sommeren, og ett kryss for resten av året aldri/ 1 pr. 2-3 pr. 1 pr. 2+ sjelden mnd. mnd. uke pr.uke Om sommeren
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (sett ett kryss) 0
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0 1 2 3-4 5-6 7+ Hvor ofte spiser du iskrem? (till dessert, Krone-is osv.) Sett ett kryss for hvor ofte du spiser iskrem om sommeren, og ett kryss for resten av året aldri/ 1 pr. 2-3 pr. 1 pr. 2+ sjelden mnd. mnd. uke pr.uke Om sommeren
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i lopet av en uke? (stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0 1 2 3-4 5-6 7+ Hvor ofte spiser du iskrem? (till dessert, Krone-is osv.) Sett ett kryss for hvor ofte du spiser iskrem om sommeren, og ett kryss for resten av året aldri/ 1 pr. 2-3 pr. 1 pr. 2+ sjelden mnd. mnd. uke pr.uke Om sommeren
Lettromme (ss)	Hvor mange egg spiser du vanligvis i lopet av en uke? (stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0
Lettromme (ss)	Hvor mange egg spiser du vanligvis i lopet av en uke? (stekte, kokte, eggerøre, omelett) (Sett ett kryss) 0
Lettromme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (sett ett kryss) 0
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (sett ett kryss) 0
Lettrømme (ss)	Hvor mange egg spiser du vanligvis i løpet av en uke? (stekte, kokte, eggerøre, omelett) (sett ett kryss) 0

Hvor ofte spiser du dessert? (Sett ett kryss pr. linje)				
aldri/ 1 pr. 2-3 pr. 1 pr. 2-3 pr. 4+ pr. uke Pudding sjokolade/karamell Riskrem, fromasj				
Hvor ofte spiser du sjokolade? (Sett ett kryss)				
aldri/ 1-3 pr. 1 pr. 2-3 pr 4-6 pr. 1+ sjelden mnd. uke uke uke pr.dag				
Mørk sjokolade				
Lys sjokolade				
Dersom du spiser sjokolade, hvor mye pleier du vanligvis å spise hver gang? Tenk deg størrelsen på en Kvikk-Lunsj sjokolade, og oppgi hvor mye du spiser i forhold til den.				
☐ 1/4 ☐ 1/2 ☐ 3/4 ☐ 1 ☐ 1,5 ☐ 2+				
Hvor ofte spiser du snacks? (Sett ett kryss) aldri/ 1-3 pr. 1 pr. 2-3 pr. 4-6 pr. 1+ pr. sjelden mnd. uke uke uke dag Potetchips				
Mølje en gang til: Dette siste spørsmålet gjelder denne "møljesesongen" Har du spist skreimølje denne vinteren? □ ja □ nei				
Hvis ja, hvor mange ganger?				
Desember Januar Februar				
Antall måltider mølje				
Dersom du spiser fiskelever, hvor mange spiseskjeer pleier du å spise hver gang? □ 1 □ 2 □ 3-4 □ 5-6 □ 7-10 □ 11 eller flere				
Bruker du da kraften/fettet som leveren er kokt i? □ ja □ nei				
Hvis ja, hvor mye bruker du hver gang? □ 1-2 ss □ 0,5 dl □ 1 dl □ mer enn 1 dl				

Appendix B

Spørreskjema start Norwegian version

Røyking & A	Alkoho	lbruk					
Røyker du?				Bruker d	u alkohol?		
□ ja □ nei				☐ ja	⊐ nei		
Hvis ja, hvor mar	nge siga	etter?	/dag		vor mange enheter i uka?		
Har du røykt mer perioder tidligere		ndre over le	engre	i gjennom	nvis det er mindre enn én enhet i uka snitt. En enhet er: øl (1/2 l), vin rennevin (drink), likør/hetvin (glass)		
fra (alder) gjennomsnittlig					Har du hatt høyere eller lavere forbruk over lengre perioder tidligere?		
fra (alder)	_ til	år,		fra (alder)	til år,		
gjennomsnittlig	siga	retter/dag		gjennoms	nittlig enheter/uke		
Hvor mange år h	ar du røy	kt daglig?	år	fra	_ til år,		
				gjennoms	nittlig enheter/uke		
Familie og s	nråkh	akarunn					
	_			Dakad at ak a			
					e snakker ulike sprak og har uppe er norsk, samisk og kvensk.		
Hvilket hjem Sett ett eller flere		råk hade	de du, dii	ne foreld	re og besteforeldre?		
Morfar Mormor Farfar Farmor Far Mor Jeg selv	Norsk	Samisk	Kvensk	Usikkert	Annet, beskriv		
Hva er din, d Sett ett eller flere		s og din	mors et	niske bal	kgrunn?		
Min etniske bakgri Fars etniske bakg Mors etniske bakg	unn er runn er		misk Kvens	k Usikkert	Annet, beskriv		
Hva regner of Sett ett eller flere		selv so	m?				
	,	Norsk	Samisk	Kvensk	Annet, beskriv □		

ID			

MS og beinskjørhet - Spørreskjema

Beinskjørhet							
Kjenner du til at beinskjørhet/ost	eoporose f	orekommer bla	nt dine foreldre	e eller søsken?			
🗖 ja 🔲 nei							
Hvis ja, hvem har beinskjørhet?_							
For kvinner							
Hvor gammel var du da du fikk m	nenstruasjo	nen for første ç	gang?	_ år			
Har du hatt regelmessig menstruasjon (d.v.s. med 3-5 ukers mellomrom) det siste året?							
☐ ja ☐ nei	•		ŕ				
Hvis du bruker p-piller eller har b	rukt p-pille	er tidligere: i hv	or mange år tils	sammen?	_ år		
Kosthold							
Har du endret kostholdet ditt ette	er at du fikk	diagnosen MS	? □ ja □ n	ei			
Hvis ja, hvordan?							
Når (årstall)?							
Har du endret kostholdet ditt av a	andre arun	ner2 🗍 ia 🔝	⊒ nei				
Hvis ja, hvordan?	andre grun	nen: 🛥 ja 🔾	■ Hei				
11115 Ju, 11101dui1.							
Når (årstall)?							
Kosthold det siste året							
I de følgende spørsmålene ønske Forsøk å tenke deg et gjennomsi			er du har brukt	det siste året.			
Hva spiser du til middag?							
	aldri	0-1 ganger i uka	2-3 ganger i uka	4-7 ganger i uka			
Fisk eller fiskemat							
Kjøtt, kjøttkaker, pølser							
Grønnsakretter Grøt, pannekaker, pizza, pasta?							
Clibit, parificitation, pizza, pasta:	_	_	_	_			
Har du gjort noen større endring	er i kosthol	ldet det siste år	et? □ ja □	nei			
Hvis ja, hvilke?							
Har du i løpet av det siste året sk	anket deg r	ner enn 5 kg ell	er torsøkt å sla	nke deg i lengre			
perioder? 🗖 ja 💢 nei							

Uteaktiviteter og soling det siste året Har du vært på solferie i utlandet etter mai 2007? Land Måned antall uker Var du i Troms eller Finnmark **hele** sommeren 2007 (juni – august)? ☐ ja ☐ nei Hvis nei, oppgi sted og varighet på reiser over en uke. Sted Antall uker Har du vært i **solarium** det siste året? ☐ nei ☐ ja, _____ (antall) ganger det siste året, hvorav _____ ganger de siste tre måneder og ___ ganger den siste måneden. Jeg soler meg i gjennomsnitt i _____ minutter hver gang. Fysisk aktivitet det siste året Er du i lønnet eller ulønnet arbeid? \Box ja ☐ nei Hvis ja, hvor mange timer i uken? Hvis du er i arbeid: Hvor stor er den fysiske belastningen? (sett ett kryss) ☐ For det meste stillesittende arbeid (for eksempel skrivebordsarbeid, montering) Arbeid som krever at du går mye (for eksempel ekspeditørarbeid, husmor, undervisning) Arbeid hvor du går og løfter mye (f.eks. postbud, pleier, bygningsarbeid) Tungt kroppsarbeid (for eksempel skogsarbeid, tungt jordbruksarbeid, tungt bygningsarbeid) Hvordan har din fysiske aktivitet i fritiden vært det siste året? Tenk deg et ukentlig gjennomsnitt for året. Arbeidsvei regnes som fritid. Besvar begge spørsmålene Timer per uke: Ingen under 1 1-2 3 og mer Lett aktivitet, ikke svett/andpusten Hard fysisk aktivitet, svett/andpusten Angi bevegelse og kroppslig anstrengelse i din fritid. Hvis aktiviteten varierer meget, så ta et gjennomsnitt. Spørsmålet gjelder bare det siste året. Sett kryss i den ruta som passer best Leser, ser på fjernsyn eller annen stillesittende beskjeftigelse.

☐ Spaserer, sykler eller beveger meg på annen måte *minst 4 timer i uka*.

Regn også med gang eller sykling til arbeidsstedet, søndagsturer m.m.

Driver mosjonsidrett, tyngre hagearbeid eller lignende.

☐ Trener hardt eller driver konkurranseidrett.

Hvor har du bodd til du	fylte 20	år?						
Hvor mange år har du bodd i Trom	s eller Finnn	nark? åı	r					
Hvor har du bodd resten av tiden?	Hvor har du bodd resten av tiden?							
Kosthold i oppveksten	(tiden før	du flyttet hj	emmefra for	godt)				
Hva spiste du til middag i oppveksten? aldri 0-1 ganger 2-3 ganger 4-7 ganger								
Fisk (kokt, stekt) eller fiskemat Kjøtt, kjøttkaker, pølser Grønnsakretter Grøt, pannekaker, pizza, pasta	_ _ _	i uka	i uka	i uka				
Hvor mye melk og yoghurt brukt (med melk menes alle melkeslag ir			.1 beger yoghu	rt tilsvarer ett glass melk)				
Gjennomsnittlig glass per da	ag (skriv "0"	hvis du ikke bru	kte melk eller y	oghurt hver dag)				
Fikk du tran i oppveksten?								
☐ ja, om vinteren (minst 4 da☐ ikke eller uregelmessig (mi	 □ ja, hele året (minst 4 dager i uka) □ ja, om vinteren (minst 4 dager i uka) □ ikke eller uregelmessig (mindre enn 4 dager i uka) 							
Har du hatt spiseforstyrrelser? User du forsøkt å slanke deg over	-	-	☐ nei					
Uteaktiviteter i oppveks	sten							
Sett kun ett kryss for hver aldersgr	uppe.							
Om sommeren: Hvor mye utendø								
Lite 6-10 år □	Midd	_	ske mye l □	Jte stort sett hele tiden				
11-15 år	Ē	_ '		ā				
16-20 år]						
Om vinteren: Hvor mye utendørsa				·				
6-10 år	Midd	_	ske mye l □	Jte stort sett hele tiden				
11-15 år		<u> </u>						
16-20 år								
Evelek aktivitet i oppveksten								
Fysisk aktivitet i oppveksten								
Angi bevegelse og kroppslig anstrengelse i <u>din fritid</u> . Her skal hele oppveksten ses under ett. Hvilket av de fire aktivitetsnivåene beskriver ditt aktivitetsnivå best?								
□ Looto oå nå flamasim sllam	hadda ======	n atilla sitta :l - !-	ookiotiaalaa					
■ Leste, så på fjernsyn eller■ Spaserte, syklet eller beve								
Regn også med gang eller syklin	g til skolen, sør	ndagsturer m.m.		-				
Drev med mosjonsidrett eller tilsvarende fysisk aktivitet.Drev med hard trening eller konkurranseidrett.								

Appendix C

Spørreskjema hver 12. uke Norwegian version

ID		

MS og beinskjørhet

Kosthold								
Har du gjort noen større	endring	er i kos	tholde	at de e	siste tre r	nånedene?		
□ ja □ nei	channy	ci i kos	morac	ot <mark>uc .</mark>		nancache.		
•								
Hvis ja, hvilke?						E ka allau f		ulca dan i
Har du i løpet av de sist	e tre man	leaene	Sianke	et deg	mer enn	i 5 kg eller i	orsøkt a sian	ike deg i
lengre perioder?								
☐ ja ☐ nei								
17 11 - 1 - 1 - 1		<u> </u>						
Kosthold den s	siste n	nane	aen					
l de følgende spørsmåle Forsøk å tenke deg et g			ar på l	nvilke	matvare	r du har bru	ıkt <mark>den siste</mark>	<mark>måneden</mark> .
Hvor ofte har du sp	ist fet fis	sk til n	nidda	ag de	n siste	måneden	?	
Sett kun et kryss på hver								
	ikke	1 gang	9	2-3 (ganger	1 pr. uke	2 pr. uke	3+ pr. uke
Laks, ørret Makrell								
Sild								
Dersom du spiser fisk, hvor mye spiser du vanligvis per gang? (1 skive/stykke = 150 gram) 1 skive								
Fiskepålegg								
Har du brukt fiskepålegg □ ja □ nei	den siste ı	månede	en?					
Hvis ja, hvor mange skiv	er med fisl					4.0	7.0	140
	0 pı	r. uke	1 pr.	uke	2-3 pr. uke	4-6 pr. uke	7-9 pr. uke	uke
Laks								
Makrell i tomat, røkt maki Sild	rell							
Kaviar								
Annet fiskepålegg								

Melk								
Har du drukket TINE ekstra lett melk (vitamin D beri	ket, grønn kartong) den s	siste måneden?						
☐ ja ☐ nei (eller mindre enn 1 glass per ul								
,	,							
Hvis ja, hvor mange glass drikker du vanligvis?								
□1-4 per uke □5-6 per uke □1 per dag □2-3 per dag □4+ per dag								
Smør og margarin								
	siata månada:-0							
Har du brukt smør eller margarin på brødskiva den s	siste maneden?							
□ ja □ nei								
Hvis ja, hvor mange skiver spiser du per dag	(antall)							
Hvor tykt lag pleier du å smøre på?								
Sett ett kryss (en kuvertpakke med smør/margarin v	eier 12 gram)							
□ skrapet (3g) □ tynt lag (5g) □ god	dt dekket (8g)	tykt lag (12g)						
	· · · · · · · · · · · · · · · · · · ·							
Tran og fiskeolje								
Har du brukt medisinsk tran eller fiskeoljekapsler den siste måneden? ja, hver dag ja, minst 4 dager i uka mindre enn 4 dager i uka nei Hvis ja, hva er navnet på tranen/fiskeoljekapslene? Mølje Har du spist skreimølje denne vinteren? ja nei Hvis ja, hvor mange ganger?								
Desember	Januar	Februar						
Antall måltider mølje								
Dersom du spiser fiskelever, hvor mange spiseskjeer pleier du å spise hver gang? 1								
Hvis ja, hvor mye bruker du hver gang? □ 1-2 ss □ 0,5 dl □ 1 dl □ mer enn 1 dl								

Soling							
Har du vært på solferie i utlandet <mark>de siste tre månedene</mark> ? □ ja □ nei							
Hvis ja, vennligst svar på følgend	de						
Sted (Land)	fra – til (dato) Omtrent hvor mange timel uke solte du deg?						
Har du vært i solarium <mark>de siste t</mark>	re måneder?						
☐ nei							
☐ ja, (antall) ganger de si	iste tre måneder, hvorav ga	nger den siste måneden. Jeg soler					
meg i gjennomsnitt i minu	tter hver gang.						
Fysisk aktivitet de	siste tre månedene						
	nstrengelse i <u>din fritid</u> . Hvis akti [,] der bare de siste tre månedene. best	viteten varierer meget, så ta et					
Leser, ser på fjernsyn ell	er annen stillesittende beskjeftigels	se.					
☐ Spaserer, sykler eller be	veger meg på annen måte <u>minst 4</u>	<u>timer i uka.</u>					
Regn også med gang eller syk	ling til arbeidsstedet, søndagsturer m.m.						
☐ Driver mosjonsidrett, tyng	gre hagearbeid eller lignende <u>mins</u>	<u>t 4 timer i uka.</u>					
☐ Trener hardt eller driver konkurranseidrett.							
Røyking							
Hoyking							
Røyker du?							
□ ja □ nei							
Hvis ja, hvor mange sigaretter?	/dag						

Har du begynt eller sluttet med medisiner de siste 3 månedene? ☐ ja nei 🗖 Angi navnet på nye medisiner du bruker (også naturmedisin, men ikke kosttilskudd) og hvilken grunn det er til at du tar disse, samt navnet på medisiner du har sluttet med. Navn på medisinen Grunn til bruk Fast Ved Begynt Sluttet (sykdom eller symptom) (dato for nye (dato) behov medisiner) Dersom det ikke er nok plass her, kan du fortsette på eget ark som du legger ved. Har du begynt eller sluttet med kosttilskudd de siste 3 måneder? ☐ ja nei 🗖 Hvis et eller flere av kosttilskuddene inneholder vitamin D, hvor mye? Noter kun navnet på kosttilskudd du har brukt daglig eller flere ganger i uka. Vitamin D begynt sluttet Grunn til bruk Navn på kosttilskuddet (dato) (dato) innhold Dersom det ikke er nok plass her, kan du fortsette på eget ark som du legger ved. For kvinner Har du hatt regelmessig menstruasjon (d.v.s. med 3-5 ukers mellomrom) de siste tre månedene? ☐ ja ☐ nei ☐ bruker hormonspiral eller p-sprøyte Kommentarer

Medikamenter og kosttilskudd



ISBN xxx-xx-xxxx-xxx-x